



Mechanisms of cannabinoid CB₂ receptor-mediated reduction of dopamine neuronal excitability in mouse ventral tegmental area

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ABSTRACT

Background: We have recently reported that activation of cannabinoid type 2 receptors (CB₂Rs) reduces dopamine (DA) neuron excitability in mouse ventral tegmental area (VTA). Here, we elucidate the underlying mechanisms.

Methods: Patch-clamp recordings were performed in mouse VTA slices and dissociated single VTA DA neurons. **Findings:** Using cell-attached recording in VTA slices, bath-application of CB₂R agonists (JWH133 or five other CB₂R agonists) significantly reduced VTA DA neuron action potential (AP) firing rate. Under the patch-clamp whole-cell recording model, JWH133 (10 μM) mildly reduced the frequency of miniature excitatory postsynaptic currents (mEPSCs) but not miniature inhibitory postsynaptic currents (mIPSCs). JWH133 also did not alter evoked EPSCs or IPSCs. In freshly dissociated VTA DA neurons, JWH133 reduced AP firing rate, delayed AP initiation and enhanced AP after-hyperpolarization. In voltage-clamp recordings, JWH133 (1 μM) enhanced M-type K⁺ currents and this effect was absent in CB₂^{-/-} mice and abolished by co-administration of a selective CB₂R antagonist (10 μM, AM630). CB₂R-mediated inhibition in VTA DA neuron firing can be mimicked by M-current opener (10 μM retigabine) and blocked by M-current blocker (30 μM XE991). In addition, enhancement of neuronal cAMP by forskolin (10 μM) reduced M-current and increased DA neuron firing rate. Finally, pharmacological block of synaptic transmission by NBQX (10 μM), D-APV (50 μM) and picrotoxin (100 μM) in VTA slices failed to prevent CB₂R-mediated inhibition, while intracellular infusion of guanosine 5'-O-2-thiodiphosphate (600 μM, GDP-β-S) through recording electrode to block postsynaptic G-protein function prevented JWH133-induced reduction in AP firing.

Interpretation: Our results suggest that CB₂Rs modulate VTA DA neuron excitability mainly through an intrinsic mechanism, including a CB₂R-mediated reduction of intracellular cAMP, and in turn enhancement of M-type K⁺ currents.

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1. Introduction

The CB₂R is a G-protein-coupled receptor that was cloned in 1993 [5]. Since then, the expression and function of CB₂Rs in the brain has

been debated. Early studies suggested that CB₂Rs were absent in the brain because CB₂ mRNA transcripts were not detected in brain tissues [1–4]. Based on these findings, the CB₂R has been considered as a “peripheral” cannabinoid receptor [5–7]. Recently, this concept has been challenged by the identification of CB₂R mRNA and receptor proteins throughout the central nervous system (CNS) [8,9], including the cerebral cortex [10], hippocampus [11], striatum [12], amygdala and brainstem [12–17]. Immunoblot and immunohistochemistry (IHC) assay also detect CB₂R-immunoreactivity or immunostaining in various

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Research in context

Evidence before this study

Cannabis sativa (marijuana) is a fibrous flowering plant that naturally produces an abundant variety of molecules, some with psychoactive effects. At least 4% of the world's adult population uses cannabis annually, making it one of the most frequently used illicit drugs in the world. The psychoactive effects of cannabis are mediated primarily through two cannabinoid receptor (CBR) subtypes, CB₁R and CB₂R. The prevailing view is that CB₁Rs are mainly expressed in central neurons while CB₂Rs are predominantly expressed in peripheral immune cells. However, this traditional view has been challenged by strong emerging evidence that shows CB₂Rs are moderately expressed and function in specific brain areas. New evidence has demonstrated that brain CB₂Rs mediate important modulations in animal cocaine-seeking behaviors (Xi et al., *Nature of Neuroscience*, Jul 24;14 [9]:1160-6, 2011), suggesting that these receptors may exist in the brain regions that regulate drug addiction. Recently, we further confirmed that functional CB₂Rs are expressed in mouse ventral tegmental area (VTA) DA neurons, and activation of VTA CB₂Rs reduces neuronal excitability and cocaine-seeking behavior (Zhang et al., *PNAS* 111 [46]:E5007-15, 2014). However, the mechanisms and the CB₂R-G protein intracellular signaling that underlies these VTA CB₂R-mediated modulations are unknown.

Added value of this study

In this paper, we elucidate the underlying mechanisms. Using cell-attached recording in VTA slices, bath-application of a CB₂R agonist (JWH133) inhibited VTA DA neuron firing in a concentration-dependent manner. In addition to JWH133, five other CB₂R agonists exhibited similar inhibition. Under the patch-clamp whole-cell recording model, JWH133 (10 μM) inhibited miniature excitatory postsynaptic currents (mEPSCs) but not miniature inhibitory postsynaptic currents (mIPSCs). However, JWH133 did not inhibit evoked EPSCs or IPSCs. In freshly dissociated VTA DA neurons, JWH133 reduced the action potential (AP) firing rate, delayed AP initiation and enhanced AP after-hyperpolarization. In voltage-clamp recordings, M-type K⁺ currents were enhanced by JWH133. This effect was absent in CB₂^{-/-} mice and abolished by co-administration of a selective CB₂R antagonist (AM630). In addition, CB₂R-mediated inhibition in VTA DA neuron firing can be mimicked by an M-current opener and blocked by an M-current blocker. Finally, CB₂R-cAMP signaling plays a role in modulating DA neuron excitability since enhancement of cAMP by forskolin reduced M-current and increased DA neuron firing rate.

Implications of all the available evidence

These results suggest that CB₂Rs modulate VTA DA neuron excitability through both synaptic and intrinsic mechanisms, including a reduction in presynaptic glutamate release and enhancement of postsynaptic neuronal M-currents. This work provides new insights into understanding the impact of brain CB₂Rs in a variety of DA system associated disease pathogenesis and potential therapeutics.

brain regions [16,18–23]. Consistent with these findings, we reported that brain CB₂Rs modulate cocaine self-administration, cocaine-enhanced locomotion and extracellular dopamine (DA) levels in the nucleus accumbens in mice [24]. This finding was further supported by a study where mice over-expressing CB₂Rs exhibit a significantly lower

level of cocaine self-administration and cocaine-enhanced locomotion compared to wild type (WT) mice [22]. Recently, we identified functional CB₂R expression in mouse VTA DA neurons, and selective activation of these CB₂Rs reduced DA neuronal excitability in vitro and in vivo preparations [25]. More recently, Stempel et al. reported that CB₂Rs mediated a cell type-specific plasticity in the hippocampal CA3 neurons [11]. However, the cellular and molecular mechanisms underlying CB₂R modulation of VTA DA neuronal excitability are poorly understood.

In the present study, we systematically investigated synaptic and intrinsic ionic channel mechanisms possibly underlying CB₂R modulation of VTA DA neurons using patch-clamp recordings. We found that activation of VTA CB₂Rs reduced the frequency of mEPSCs but not mIPSCs in DA neurons. By using a perforated patch-clamp recording in acutely dissociated VTA DA neurons, a simple single cell model without intact pre-synaptic connections, we found that JWH133 reduced DA neuron action potential (AP) firing rate, prolonged AP initiation, enhanced after-hyperpolarization, and enhanced M-type K⁺ channel currents. In addition, we determined whether the presynaptic (reduction of mEPSC frequency) or postsynaptic (intrinsic) or both contribute to JWH133-induced reduction in AP firing, and found that the intrinsic mechanism such as enhancement of M-current, contributes to CB₂R-mediated reduction of VTA DA neuronal excitability, which may play an important role in the regulation of DA-associated behaviors and diseases.

2. Methods and materials

2.1. Animals

Male wild-type (WT) and Zimmer strain CB₂^{-/-} mice [6,7] with C57BL/6J genetic backgrounds were bred at Barrow Neurological Institute. Genotyping was performed in our laboratory before experiments were begun. All experiments were in compliance with the Guide for the Care and Use of Laboratory Animals and experimental protocol has been approved by IACUC of Barrow Neurological Institute. When we tested the effects of CB₂R agonist on VTA DA neurons using CB₂^{-/-} mice, the littermate WT mice were used as the control. All experimental procedures were approved by the Institutional Animal Care and Use Committee at the Barrow Neurological Institute and by the Animal Ethics Committee of Qingdao University.

2.2. VTA slice preparation

Mice (14–31 days old) were used as previously described [26–28]. Briefly, mice were housed under standard conditions at 22–24 °C and 50–60% humidity under a 12 h light/dark cycle. On the experimental day, mice were anesthetized (isoflurane USP) and then sacrificed by decapitation. Horizontal (or coronal) midbrain slices (250 μm) were cut using a vibratome 1000 (Vibratome 1000 plus; Jed Pella Inc., Redding, CA) to include the VTA. Slices were prepared in an ice-cold, sucrose-based, artificial cerebrospinal fluid containing (in mM): 212.7 sucrose, 5 KCl, 1.25 NaH₂PO₄, 3 MgCl₂, 1 CaCl₂, 26 NaHCO₃, and 10 glucose. The slices were then incubated for at least 1 h in a pre-incubation chamber (Warner Ins., Holliston, MA) at room temperature (22–24 °C) in conventional artificial cerebrospinal fluid containing (ACSF, in mM): 124 NaCl, 5 KCl, 2 CaCl₂, 2 MgCl₂, 1.25 NaH₂PO₄, 26 NaHCO₃ and 10 glucose. The ACSF was continuously saturated with 95% O₂ and 5% CO₂.

2.3. Cell-attached patch-clamp recordings in VTA slices

For slice patch recordings, horizontal or coronal midbrain slices (250 μm thickness) were prepared from 14- to 31-day old mice (WT and CB₂^{-/-}) mice as described previously [28]. To maintain an intact intracellular environment, we employed a cell-attached mode of patch recording. Thus, we were able to measure stable extracellular action

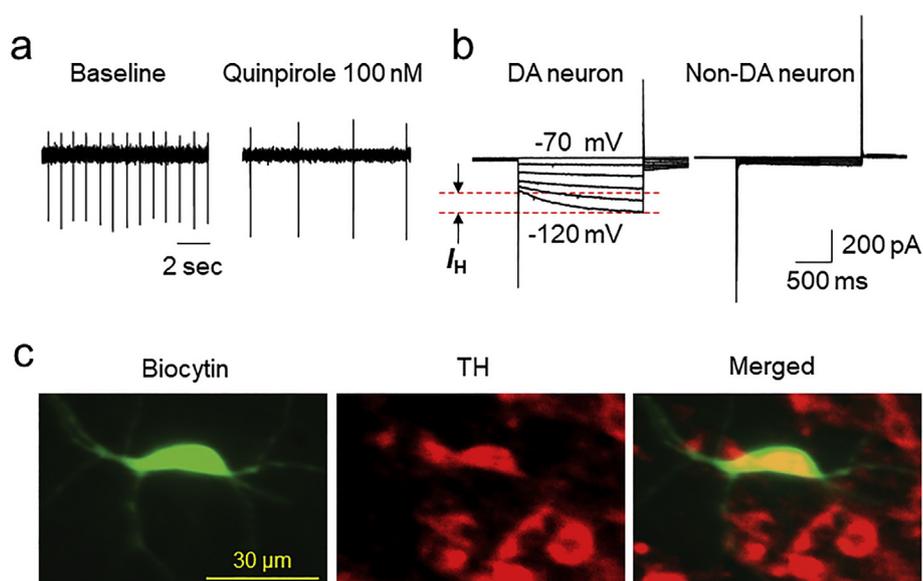


Fig. 1. Identification of VTA DA neurons from patch-clamp recording in VTA slice. (a) In patch-clamp cell-attached recording mode, bath-application of 100 nM quinpirole (dopamine D_2 receptor agonist) inhibited firing rate of DAergic neurons. (b) Under whole-cell recording mode, DA neurons exhibited hyperpolarizing pulse-activated currents (H-current, left traces) whereas GABAergic neurons usually did not express H-currents (Right traces). (c) After patch recording, the biocytin (2 mg/ml) was delivered into recorded neuron (left panel). Tyrosine hydroxylase (TH) staining was performed post hoc and labels DA neurons (middle and right panels).

potential (AP) firing for hours. The recording pipette was filled with intracellular solution containing in mM: 140 potassium gluconate, 5 KCl, 10 HEPES, 0.2 EGTA, 2 $MgCl_2$, 4 MgATP, 0.3 Na_2GTP and 10 Na_2 -phosphocreatine (pH 7.3 with KOH). After patch recording, we converted from cell-attached mode to conventional whole-cell recording to identify the neuronal phenotype of the recorded neuron according to standard electrophysiological, pharmacological and immunostaining criteria as described previously [28,29]. Briefly, in this study, we mostly recorded DA neurons from lateral VTA, which demonstrated a low action potential (AP) firing rate (1–3 Hz), large H-current, and TH positive reaction by immunostaining (Fig. 1A–C). We have analyzed 12 VTA neurons (from 12 mice), under the cell-attached recording mode to record extracellular AP firing, the averaged AP duration was 2.34 ± 0.07 ms, the averaged firing rate was 1.65 ± 0.14 Hz, and after perfusion of 100 nM quinpirole for 10 min, the AP firing rate was reduced to 0.53 ± 0.08 Hz (paired t -test $p < 0.0001$). After washout of quinpirole for 10 min, AP firing rate recovered to 1.40 ± 0.12 Hz (quinpirole washout $p < 0.0001$). In addition, we have examined the effects of JWH133 on H-currents of DA neurons in mouse VTA slices. In 4 cells tested (from 2 mice), the amplitudes of H-currents (hyperpolarizing from -60 to -120 mV) before and after exposure of 10 μM JWH133 for 10 min were 166.0 ± 24.3 pA and 161.3 ± 11.2 pA (paired t -test, $p = 0.86$). These results suggest that acute application of 10 μM JWH133 to VTA slices did not alter H-current.

2.4. Miniature or evoked excitatory/inhibitory postsynaptic currents (mEPSCs/mIPSCs, eEPSCs/eIPSCs) in VTA slices

To determine whether brain CB_2 R modulate presynaptic glutamate or GABA release, thereby altering postsynaptic DA neuronal activity, we recorded mEPSCs in the presence of TTX (a voltage-dependent Na^+ channel blocker, 1 μM), D-APV (a NMDA receptor antagonist, 50 μM) and picrotoxin (a selective $GABA_A$ receptor antagonist, 100 μM), or mIPSCs in the presence of TTX, NBQX (an AMPA receptor antagonist, 10 μM) and D-APV (50 μM) in VTA DA neurons. In addition, we recorded electrical stimulation-evoked EPSCs (eEPSCs) or -evoked IPSCs (eIPSCs) in the absence of TTX to determine whether JWH133 alters eEPSCs or eIPSCs in VTA DA neurons.

2.5. Patch-clamp recording from single dissociated VTA DA neurons

Perforated-patch whole-cell recording techniques were employed [30,31]. Compared to performing conventional whole-cell recordings, this approach was crucial to obtain stable nicotinic responses from dissociated VTA neurons, presumably due to minimal perturbation of the intracellular environment. Pipettes (3–5 $M\Omega$) used for perforated-patch recordings were filled with intracellular recording solution, which contained (in mM): 140 K-gluconate, 10 KCl, 5 $MgCl_2$, and 10 HEPES; pH 7.2 (with Tris-OH) freshly supplemented before use with amphotericin B to 200 $\mu g/mL$ from a 40 mg/mL DMSO stock. The liquid junction potential was 14 mV and calculated using Clampex 9.2 (Axon Instruments, Foster City, CA), and corrections were made for junction potentials post-hoc. After tight seal (>2 G Ω) formation, about 5–20 min was required for conversion to the perforated-patch mode, and an access resistance of <60 $M\Omega$ was accepted to start the experiments. Series resistance was not compensated in this study. Data were acquired at 10 kHz, filtered at 2 kHz, digitized on-line (Digidata 1440 series A/D board, Axon Instruments), and displayed and stored on a PC computer. Rapid application of drugs was performed using a computer-controlled “U-tube” system [30]. Periods of drug exposure are specified in the text and figures or legends. All experiments were performed at room temperature (21 ± 1 $^{\circ}C$). To enable identification of single, dissociated VTA neurons, cells were labeled with biocytin (Sigma-Aldrich, St. Louis, MO; 2.0 mg/mL in the recording electrode) in some experiments. After conversion from perforated to a conventional whole-cell recording mode, the dye was injected into the cytoplasm by a pulse (200 ms, 0.5 Hz) of depolarizing current (1.0 nA) for 3 min. Labeled cells were visualized using epifluorescence microscopy (see detail in our recent publication) [25].

2.6. Data analyses and statistics

All data are presented as means (\pm S.E.M.). One-way or two-way analysis of variance (ANOVA) was used to analyze the significance of the effects of JWH133 or other drugs on neuronal firing. Individual group comparisons were carried out using the Student Newman-Keuls or Tukey method. In addition, paired (or unpaired) t -tests were also

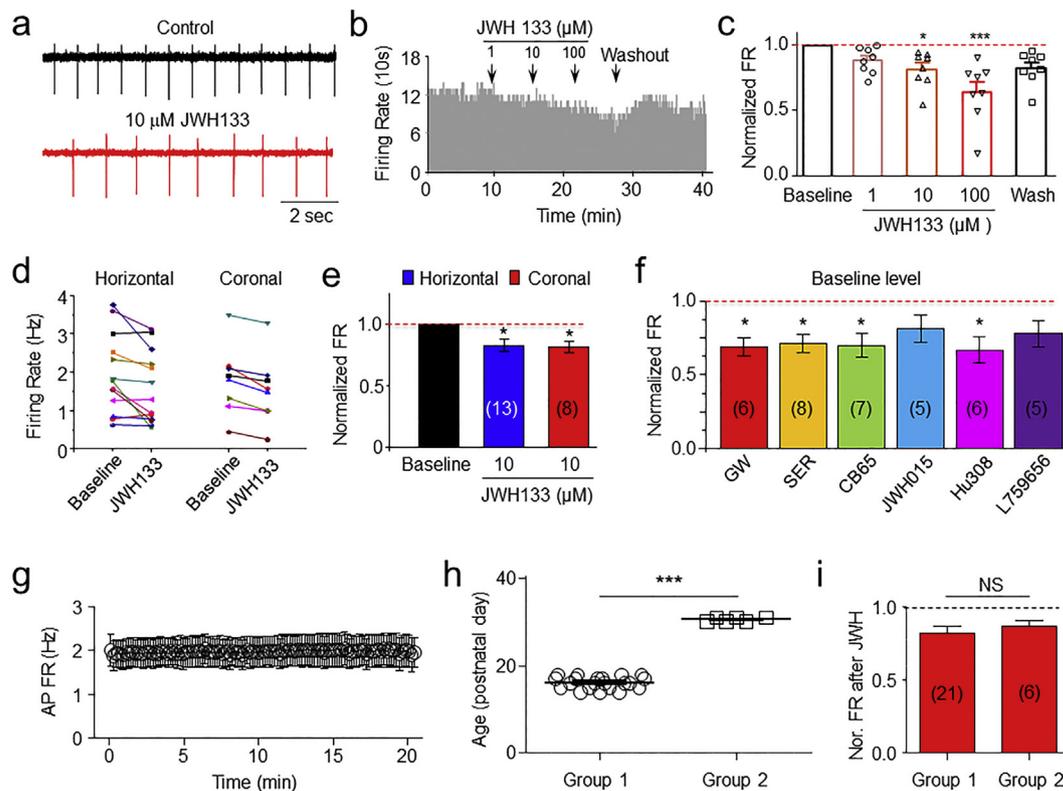


Fig. 2. Activation of VTA CB₂R reduced VTA DA neuron firing rate. In VTA slices, cell-attached recording showed the typical traces of extracellular action potential (AP) firing before and after JWH133 exposure (a). A time-frequency histogram (b) and Bar graph from the normalized data (c) showed that JWH133 inhibited VTA DA neuron firing in a concentration-dependent manner. (d) Comparison of the effects of JWH133 on AP firing rate between horizontal and coronal VTA slices, and showed no significant difference between these two types of slices (e). (f) In addition to JWH133, other 6 CB₂R agonists tested showed 4 of them exhibited inhibitory effects on VTA DA neurons. * $p < 0.05$, ** $p < 0.01$. (g) In control experiments (without any drugs), cell-attached patch recording showed a stable AP firing of VTA DA neurons for 20 min. (h) Mice were classified into two age groups (postnatal 16 and 30). (i) Statistical comparison of JWH133 (10 μ M)-induced inhibition of VTA DA neuron firing rate between these two age-groups, and showed no statistical significance (N.S.).

used to analyze some electrophysiological data as described previously [30,31].

3. Results

3.1. Activation of CB₂R reduced VTA DA neuron firing

Initial experiments were designed to examine the concentration-dependent effect of CB₂R agonist (JWH133) on VTA DA neuron firing using cell-attached recording mode. Fig. 2A shows a typical trace of VTA DA firing before (A top trace) and during (A bottom trace) CB₂R agonist JWH133 exposure. Bath applications of different concentrations of JWH133 (1, 10, 100 μ M) reduced DA neuron firing rate in a concentration-dependent manner (Fig. 2B, C, One-way ANOVA, $F_{[4,35]} = 6.97$, $p < 0.001$). Then, we examined the effects of JWH133 on DA neuron firing from horizontal ($n = 13$ from 4 mice) or coronal ($n = 8$ from 3 mice) VTA slices, and found a modest but significantly consistent effect on inhibition of VTA DA neuron firing in both types of slice preparations (One-way ANOVA, $F_{[2,31]} = 4.79$, $p < 0.05$, Fig. 2D, E). Further, Tukey analysis showed that compared to baseline, JWH133 inhibited DA firing rate in either coronal ($p < 0.05$, $n = 8$, Tukey analysis) or horizontal ($p < 0.05$, $n = 13$, Tukey analysis) slices. Furthermore, we tested whether different CB₂R agonists exhibit similar inhibitory effects on VTA DA neuron firing. As shown in Fig. 2F, the CB₂R agonists, GW, SER, CB65, JWH105, Hu308 and L759656 exhibited a similar reduction of DA firing rate (One-way ANOVA, $F_{[6,31]} = 2.4$, $p < 0.05$), but Tukey analysis showed that JWH105 and L759656 did not reach statistical significance ($p > 0.05$, Tukey analysis). Finally, to exclude the possibility that JWH133-induced reduction of AP firing rate was mediated by AP firing

rundown with time, we performed a group of control experiments, in which, VTA DA neuronal AP firing was recorded using cell-attached patch for 20 min without any drugs. In 6 slices tested (from 3 mice), there was no functional rundown of AP firing within 20 min (Fig. 2G), demonstrating that cell-attached recording to measure extracellular AP firing was stable. Considering the ages of mice used in this study were between 2 and 4 weeks, we classified mice into two age groups and compared the effects of JWH133. Group one contained 21 slices with an average of age 16.1 ± 0.3 postnatal days and group two contained 6 slices with an average of 30.5 ± 0.6 postnatal days. The difference of the ages between these two groups were statistically significant ($p < 0.001$, unpaired t -test, Fig. 2H). 10 μ M JWH133 inhibited VTA DA AP firing rate in a similar rate (Fig. 2I), in group one, the inhibitory rate was $18 \pm 4\%$ ($n = 21$ from 8 mice), while in group two, the inhibitory rate was $15 \pm 3\%$ ($n = 6$ from 2 mice, $p = 0.59$, unpaired t -test), suggesting that during animal developing between P16 and P30, the CB₂R-mediated inhibition in VTA DA neuronal firing rate is not changed. Together, these results are further consistent with our previous findings that activation of VTA CB₂R reduces VTA DA neuron firing rate [25].

3.2. Effects of CB₂R activation on excitatory synaptic transmission in VTA DA neurons

VTA DA neuron firing is controlled by both presynaptic and postsynaptic mechanisms. To elucidate the mechanisms underlying CB₂R-mediated reduction of VTA DA neuronal firing, we tested the effects of CB₂R agonist on excitatory synaptic transmission measured by miniature excitatory postsynaptic currents (mEPSCs) in DA neurons of VTA slices. The identification of VTA DA neurons was based on their

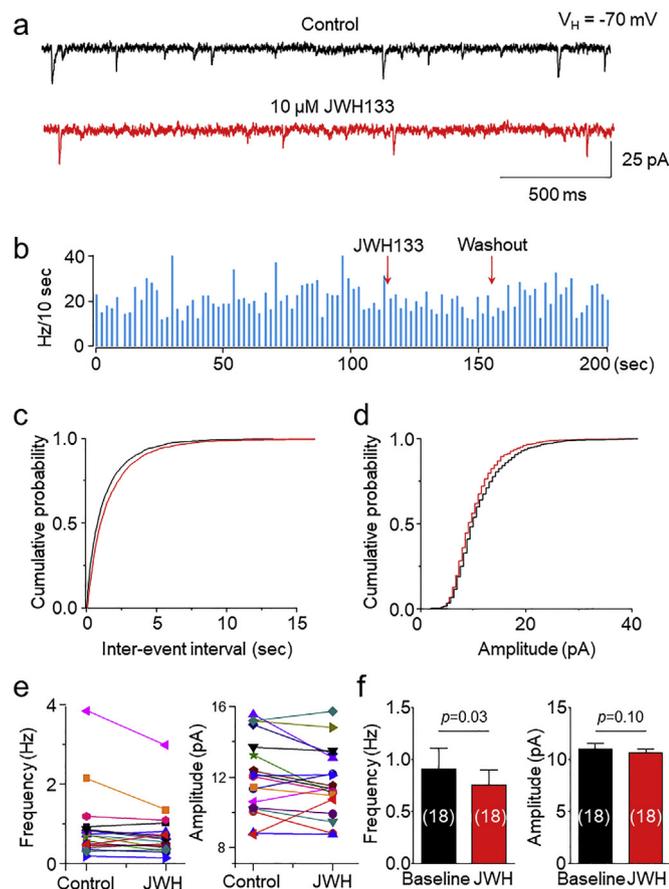


Fig. 3. Effects of JWH133 on excitatory synaptic transmission in VTA DA neurons. (a) Typical traces of mEPSCs in the presence of TTX (1 μ M) and picrotoxin (100 μ M) before (top trace) and after (bottom trace) JWH133 (10 μ M) exposure using whole-cell patch-clamp recording in a VTA slice. In 19 slices, cumulative probability analysis for mEPSC inter-event interval (b) and amplitude (c) under control conditions (black curve) and JWH133 exposure (red curve). (d) Comparison of mEPSC frequency (left panel) and amplitude (right panel) before and after JWH133 exposure from 19 VTA slices tested. (e) Bar graph data illustrating JWH133 significantly reduced mEPSC frequency, but not amplitude.

electrophysiological characteristic features and the TH staining after patch-clamp recording as described previously [25,28] and shown in Fig. 1A–C. In 18 VTA slices tested prepared from 9 WT mice, bath perfusion of 10 μ M JWH133 for 10 min altered mEPSCs (Fig. 3A,B), represented as reduced frequency in 14 slices (Fig. 3C,E, left panel) and enhanced frequency in 4 slices. Averaged data showed that JWH133 reduced mEPSC frequency from 0.91 ± 0.19 to 0.75 ± 0.14 Hz ($p = 0.03$, paired t -test, $n = 18$, Fig. 3F, left panel). Further cumulative distribution of mEPSC inter-event interval showed significantly shift ($K-S$ test, $p = 0.002$). For mEPSC amplitude, JWH133 reduced amplitude in 3 slices and enhanced amplitude in 14 slices; and no change in 1 slice (Fig. 3D, E, right panel), and together, there was no significant change of amplitude (from 11.0 ± 0.5 to 10.6 ± 0.4 pA, $p = 0.10$, paired t -test, $n = 18$, Fig. 3F, right panel). These results suggest that acute activation of CB_2 R shows a modest, but statistically significant, reduction in glutamatergic synaptic transmission in VTA DA neurons, including reduced presynaptic glutamate release probability.

3.3. Effects of CB_2 R activation on inhibitory synaptic transmission in VTA DA neurons

We then examined the effects of JWH133 on miniature inhibitory postsynaptic currents (mIPSCs) in the presence of TTX (1 μ M), NBQX (10 μ M) and D-APV (50 μ M). In 23 VTA slices prepared from 11 WT

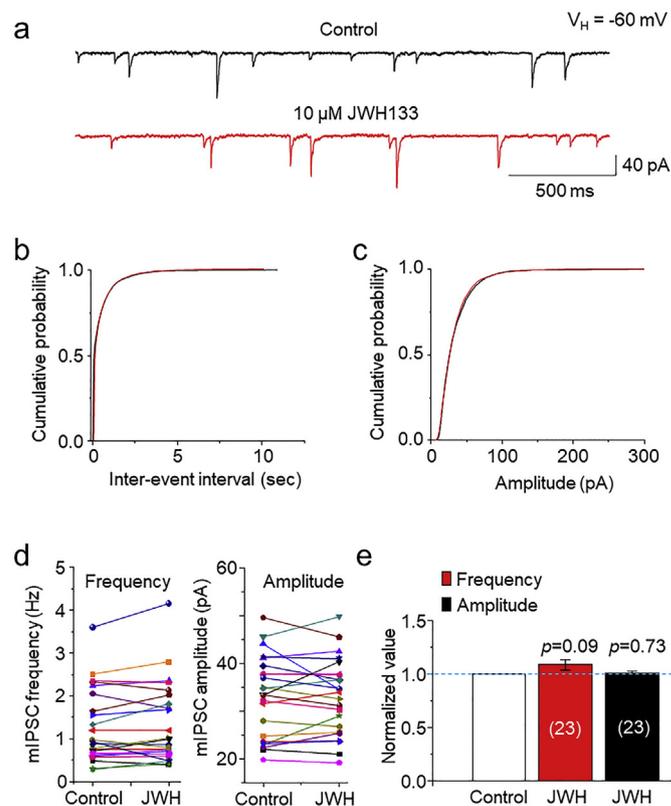


Fig. 4. Effects of JWH133 on inhibitory synaptic transmission in VTA DA neurons. (a) Typical traces of mIPSCs in the presence of NBQX (10 μ M) and D-APV (50 μ M) before (top trace) and after (bottom trace) JWH133 (10 μ M) exposure using the whole-cell patch-clamp recording in a VTA slice. (b) and (c) In 23 VTA slices, cumulative probability analysis for mIPSC inter-event interval and amplitude under control conditions (baseline) and JWH133 exposure. $K-S$ Test showed that JWH133 did not significantly alter both mIPSC frequency ($p = 0.432$, b) and amplitude ($p = 0.384$, c). (d) Comparison of mIPSC frequency (left panel) and amplitude (right panel) before and after JWH133 exposure from 23 VTA slices tested. (e) Normalized bar graph data illustrating JWH133 did not alter mIPSC frequency and amplitude.

mice, bath perfusion of 10 μ M JWH133 for 10 min altered mIPSC frequency and amplitude, represented as reduced frequency in 9 slices and enhanced frequency in 14 slices; reduced amplitude in 8 slices and enhanced amplitude in 14 slices; and no change in one slice. After averaged, these data showed that after JWH133 exposure, mIPSC frequency was 109% ($109 \pm 5\%$, $p = 0.09$, paired t -test, $n = 23$) and amplitude was 101% ($101 \pm 2\%$, $p = 0.73$, paired t -test, $n = 23$) compared to the baseline level (Fig. 4A, E). Neither inter-event interval nor amplitude cumulative distribution showed shift (Fig. 4B, C). These results suggest that activation of CB_2 R exhibits little effects on either presynaptic GABA release or postsynaptic GABA_A receptor function.

3.4. Effects of CB_2 R activation on evoked EPSCs and IPSCs in VTA DA neurons

We also examined the effects of JWH133 on evoked EPSCs (eEPSCs) and IPSCs (eIPSCs). In these experiments, 100 μ M PTX (for eEPSCs) or NBQX (10 μ M) + APV (50 μ M) (for eIPSCs) was added into the bath solution. A bipolar tungsten stimulation electrode was placed at ~ 150 μ m rostral to the recording electrode and the paired stimulation pulses with 50 ms interval were applied (Fig. 5A). The results showed that bath perfusion of 10 μ M JWH133 for 10 min failed to alter the current amplitude and the P2/P1 ratio of paired-pulse-induced eEPSCs (Fig. 5A–C) and eIPSCs (Fig. 5D–F). These results suggest that acute activation of VTA CB_2 R does not affect evoked synaptic neurotransmitter release.

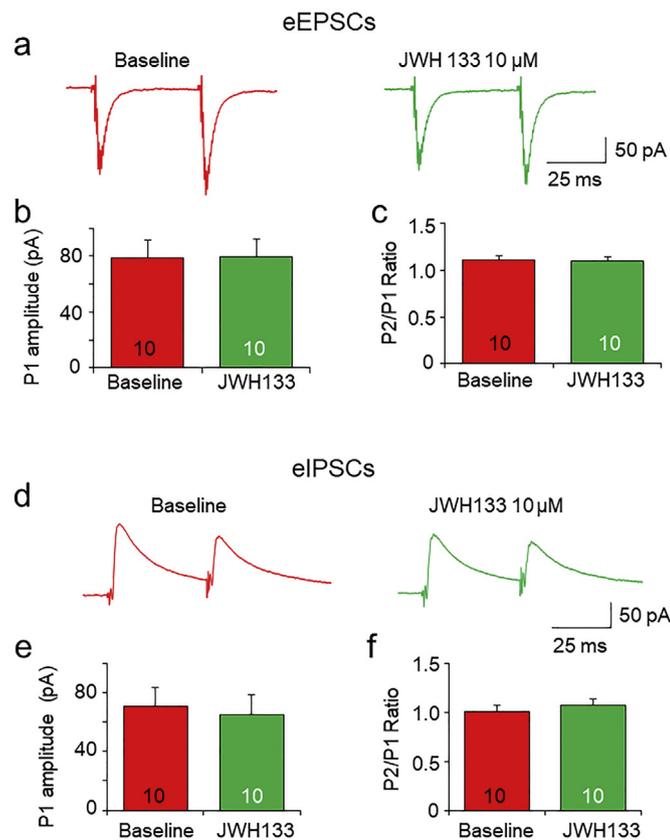


Fig. 5. Effects of JWH133 on evoked EPSCs and IPSCs in VTA DA neurons. (a) Representative typical traces of paired-pulse (50 ms interval) stimulation-induced eEPSCs before (left trace pair) and after (right trace pair) exposure to JWH133. Bar graph illustrating that JWH133 (10 μ M) altered neither eEPSC amplitude (b) nor P2/P1 ratio (c) in 10 VTA slices tested. (d) Representative typical traces of paired-pulse (50 ms interval) stimulation-induced eIPSCs before (left trace pair) and after (right trace pair) exposure to JWH133. Bar graph illustrating that JWH133 (10 μ M) did not change eIPSC amplitude (e) and P2/P1 ratio (f) in 10 VTA slices tested.

3.5. Characteristic features of CB₂R-mediated modulation in VTA DA neuron action potentials

The data presented thus far demonstrated that a mild reduction of presynaptic glutamatergic by acute CB₂R activation. We further elucidated possible intrinsic mechanisms of the CB₂R-mediated modulation in postsynaptic DA neurons. To accomplish this, we used freshly dissociated single VTA DA neurons, which is a pure postsynaptic cell model since the enzymatic treatment and mechanical dissociation remove all presynaptic terminals/boutons from these cells [25,30,31]. Perforated (amphotericin B) patch-clamp recording from the single dissociated VTA DA neurons showed that bath application of 1 μ M JWH133 for 1–3 min reduced AP firing rate (Fig. 6A–C), prolonged AP initiation (Fig. 6D) and enhanced the amplitude of after-hyperpolarization current (Fig. 6E). Fig. 6F summarizes these AP alterations before, during and after exposure to JWH133. These results suggest that, in a single neuron model, the activation of somatodendritic CB₂R in VTA DA neurons reduces neuronal excitability likely via alterations of intrinsic ion channels such as the M-type K⁺ channels.

3.6. Effects of CB₂R agonist on voltage-dependent M-type K⁺ currents

M-channel is a voltage-gated, non-inactivating K⁺ channel. It is expressed in VTA DA neurons [32] and functionally modulates neuronal excitability [33,34]. It has been reported that M-channels are modulated by multiple G-protein-coupled receptors, including CB₁R [35].

Therefore, we explored the possibility that CB₂R may also modulate M-currents in VTA DA neurons. M-currents were induced with a standard deactivation voltage protocol [32,36] as shown in Fig. 7A. M-current is measured as the inward relaxation current caused by M-current deactivation during the voltage steps. Bath-applied JWH133 (1 μ M) significantly enhanced M-currents elicited by three hyperpolarizing holding potentials in dissociated VTA DA neurons in WT mice (Fig. 7B), and this effect was reversible after JWH133 washout (Fig. 7C). Fig. 7D shows the superimposed traces of M-currents before and during JWH133 exposure, showing a JWH133-induced M-current augmentation. This effect was blocked by AM630 (1 μ M) and absent in VTA DA neurons prepared from CB₂^{-/-} mice. To confirm the identity of the recorded M-current, we examined the effects of retigabine (a selective M-current enhancer, 10 μ M) and XE991 (a selective M-current blocker, 30 μ M) on the recorded current. Like JWH133, retigabine enhanced ($n = 5$ from 3 mice, $p = 0.02$, paired t -test), but XE991 reduced ($n = 8$ from 4 mice, $p = 0.003$, paired t -test) the recorded currents produced by the M-current-inducing protocol, supporting the conclusion that the recorded currents were, indeed, M-currents. Fig. 7E shows pooled data, illustrating the effects of each individual group tested, in which JWH133 (1 μ M) enhanced M-currents ($n = 19$ from 8 mice, $p = 0.0002$, paired t -test), AM630 + JWH133 abolished JWH133's effect, retigabine enhanced but XE991 reduced M-currents in WT mice. While in CB₂^{-/-} mice, JWH133 failed to enhance M-current but retigabine still enhances and XE991 still reduced M-currents. These results suggest that the inhibitory effect of JWH133 on M-current is mediated through the CB₂R, while the effects of retigabine and XE991 on M-current are CB₂R-independent.

3.7. Roles of CB₂R-mediated enhancement of M-current in altered VTA DA neuron firing

To further determine whether JWH133-enhanced M-currents contribute to JWH133-induced VTA DA neuronal inhibition, we examined the effects of retigabine and XE991 on VTA DA neuronal firing. In single dissociated VTA DA neurons, we found that retigabine (10 μ M) increased M-currents, in an amplitude similar to JWH133, causing a reduction in VTA DA neuronal firing, while XE991 (30 μ M) attenuated M-currents, producing an increase in neuronal firing (Fig. 8A, B). Fig. 8C and D show the summarized results after statistical analyses that retigabine reduced AP firing rate and prolonged AP initial time, and XE991 increased AP firing rate and shortened AP initial time. These results suggest that M-current in VTA DA neurons play an important role in modulation of neuronal firing rate. In VTA slices, cell-attached recordings showed that, in the presence of XE991, JWH133 failed to inhibit DA neuron firing (Fig. 8E, F). These results suggest that JWH133-enhanced M-currents play a role in the JWH133-induced reduction in VTA DA neuronal firing.

3.8. Intracellular signal pathway of JWH133-induced enhancement of M-current

In these experiments, we determined which second messenger molecule mediates the CB₂R-G-protein signaling to M-type K⁺ channels. It is known that CB₂R activation primarily initiates two signal pathways, MAPK/ERK and cAMP/PKA, producing an increase in the former and decrease in the latter [37–39]. Since cAMP, but not MAPK, is associated to M-type K⁺ channels [37,40], we examined the effect of enhanced intracellular cAMP by forskolin on JWH133-induced enhancement of M-current. As shown in Fig. 9, bath-perfusion of 10 μ M forskolin to dissociated single VTA DA neuron reduced M-current by $19.2 \pm 4.9\%$ and in the presence of forskolin, JWH133 (1 μ M) potentiated M-current by $8.1 \pm 3.7\%$ ($n = 10$ from 5 mice, One-Way ANOVA, $F_{[3,27]} = 14.9$, $p < 0.0001$). Further analysis showed that forskolin-induced reduction of M-current was significant ($p < 0.01$, One-way ANOVA, Tukey), while in the presence of forskolin, JWH133-induced change of AP firing was

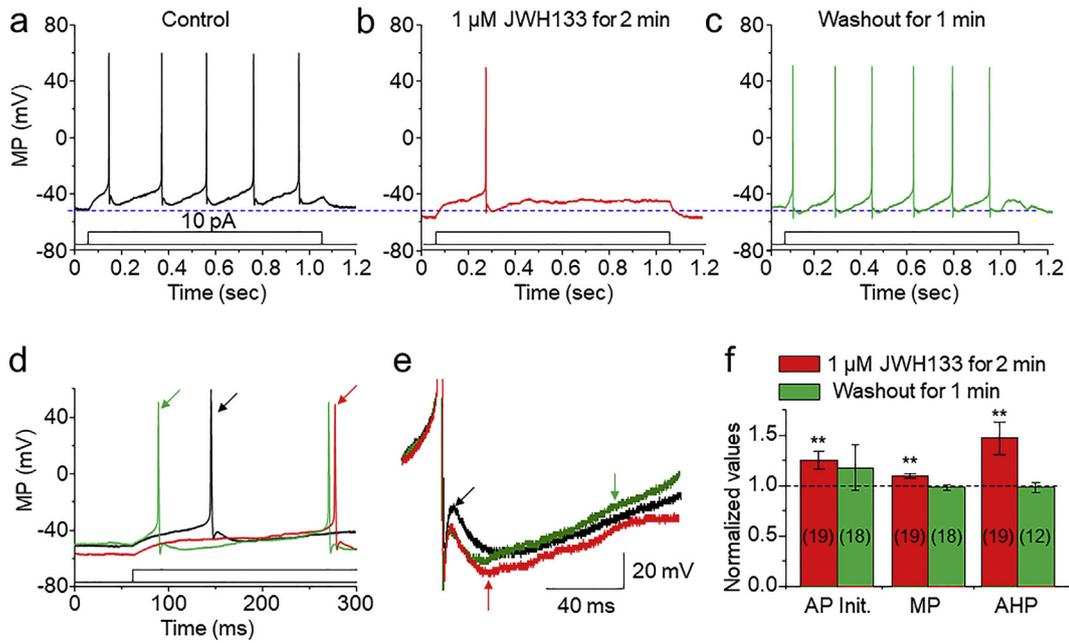


Fig. 6. Analysis of CB₂R-mediated changes of action potential (AP) in dissociated VTA DA neurons. Representative typical traces of AP of VTA DA neurons before (a), during (b) and after (c) exposure of 1 μM JWH133 under perforated current-clamp recording conditions. AP firing was elicited by injection of 10 pA current through recording electrode. Superimposed the first AP from trace (a)-(c) to show the AP initiation (d) and AP after-hyperpolarization (e) with expended time scale. (f) Normalized data showing the effect of JWH133 on AP firing parameters. * *p* < 0.05, ** *p* < 0.01.

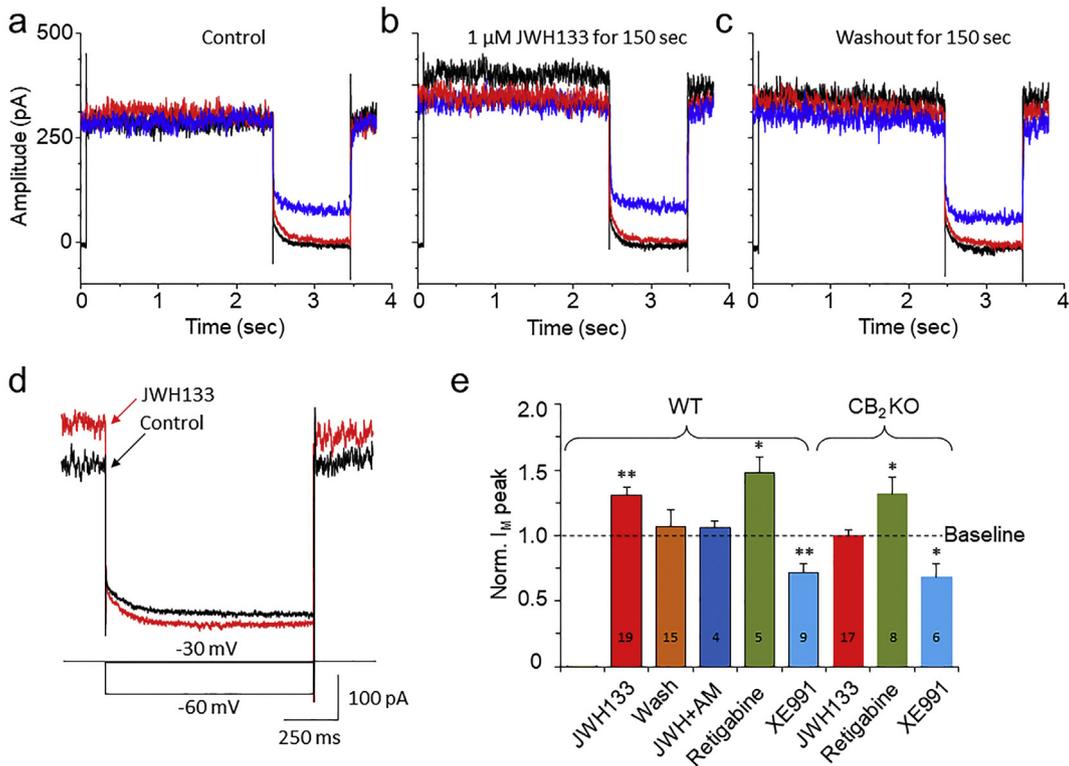


Fig. 7. Effects of JWH133 on M-type K⁺ currents in single dissociated VTA DA neurons. (a)-(c) Representative typical traces of M-currents before, during and after exposure of JWH133. M-currents were induced by a pre-depolarization from holding potential of -60 mV to -30 mV, then three steps back to -60 mV in 10 mV increments. Fig. 6b illustrates that JWH133 increased M-current amplitude in VTA DA neurons. (d) Superimposed M-current before (Black trace) and during (red trace) JWH133 exposure with expended time scale. (e) Bar graph illustrating that JWH133 (1 μM) enhanced M-currents, which was blocked by AM630 or absent in CB₂^{-/-} mice. Retigabine enhanced, while XE991 attenuated M-currents in both WT and CB₂^{-/-} mice. The above experiments were performed separately by each paired (before and after tested drug exposure) group. In statistical analyses, the baseline (before drug exposure) was normalized as 1, and the changed value after drug perfusion was compared to the baseline of each individual group, and analyzed using paired *t*-test.

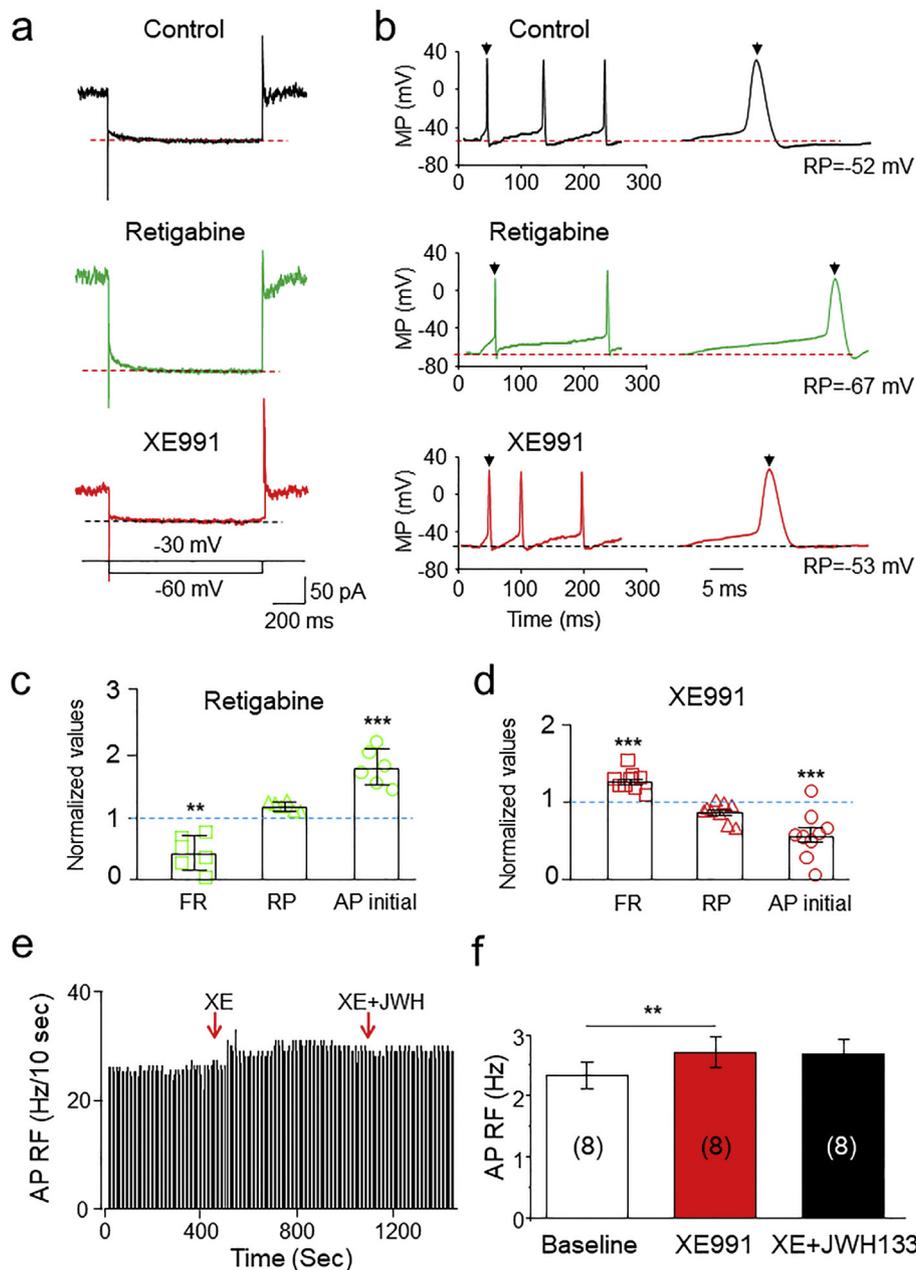


Fig. 8. Roles of M-type K^+ currents in JWH133-modulated VTA DA neuron firing. (a) Representative typical traces of M-type K^+ currents in control (top trace), retigabine (middle trace) and XE991 (bottom trace) from the same acutely-dissociated neuron, and showed that retigabine enhanced and XE991 reduced M-current. (b) Representative typical traces of AP firing in control (top trace), retigabine (middle trace) and XE991 (bottom trace) from the same recorded neuron, and showed that retigabine decreased and XE991 increased AP firing rate. (c) Statistical analysis of the effects of retigabine on VTA DA neuronal AP firing including firing rate (FR), resting membrane potential (RP) and AP initial time. AP firing was evoked by an injection of depolarizing current (20 pA). (d) Statistical analysis of the effects of XE991 on VTA DA neuronal AP firing including firing rate (FR), resting membrane potential (RP) and AP initial time. AP firing was evoked by an injection of depolarizing current (20 pA). (e) In VTA slices using cell-attached recording, the time-frequency histogram showed no detectable change of AP firing rate after exposure to XE991 plus JWH133. (f) Statistical analysis showed JWH133 failed to reduce VTA DA firing rate in the presence of XE991.

not significant ($p > 0.05$, Fig. 9A,B, One-way ANOVA, Tukey). These results suggest that CB_2R -mediated reduction of intracellular cAMP likely contributes to the enhancement of M-current by JWH133. Finally, we determined whether forskolin can prevent the JWH133-induced DA firing reduction. As shown in Fig. 9C and D, using cell-attached recording in VTA slices, bath-application of forskolin (10 μ M) increased DA firing rate and prevented JWH133 (10 μ M)-mediated reduction of DA firing rate ($n = 11$ from 5 mice, One-way ANOVA, $F_{[2,20]} = 4.63$, $p = 0.018$, Fig. 9C, D). Further analysis showed that forskolin increased AP firing rate by $8.2 \pm 2.0\%$ ($p < 0.05$, paired t -test), while in the presence of forskolin, JWH133 altered AP firing by $1.2 \pm 3.0\%$ ($p > 0.05$, paired t -test). Together, these results suggest that JWH133 activates CB_2R -G-protein signaling, thereby reducing intracellular cAMP levels, in turn

enhancing M-type K^+ channel function, and ultimately reducing VTA DA neuronal excitability.

3.9. Do both synaptic and intrinsic mechanisms contribute to CB_2R -mediated reduction of VTA DA neuron firing?

To further determine the above findings that presynaptic mechanism (reduced mEPSC frequency) and intrinsic mechanism (enhanced M-current) contribute to CB_2R -mediated inhibition in VTA DA neuronal firing, we designed two additional experiments. First, we examined the effects of JWH133 on neuronal firing in the presence of NBQX (10 μ M), APV (50 μ M) and picrotoxin (100 μ M) to block excitatory and inhibitory presynaptic transmissions. We found that under cell-attached recording

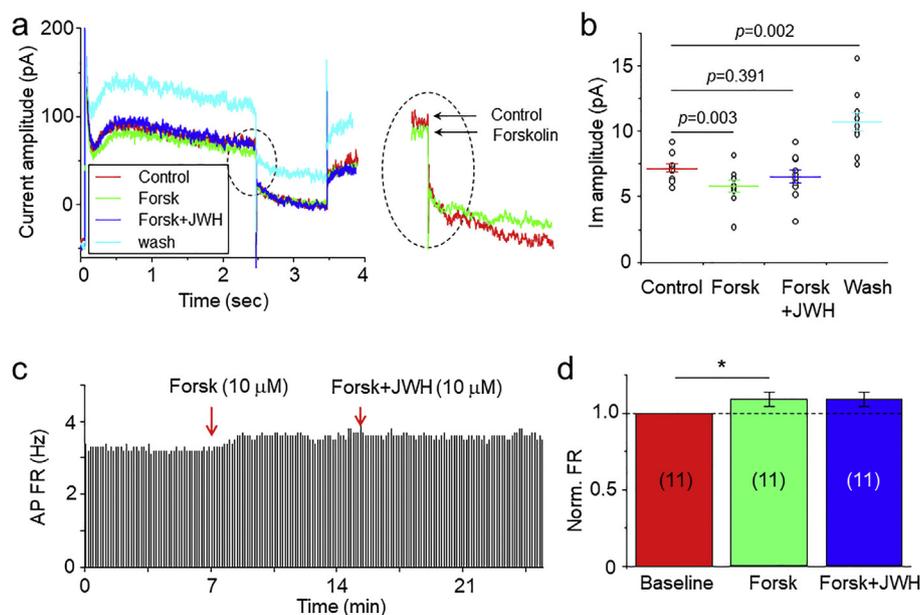


Fig. 9. Effects of forskolin on JWH133 modulated M-current and AP firing in VTA DA neurons. (a) Representative the typical traces of M-current under four different conditions: control (control, red trace), forskolin (Forsk, green trace), forskolin plus JWH133 (blue trace) and washout of drugs (light blue trace). The enhanced inset showed a reduction of M-current by forskolin. (b) Statistical analysis showed that forskolin significantly reduced M-current and in the presence of forskolin, JWH133 failed to enhance M-current. (c) In VTA slices using cell-attached recording, a time-frequency histogram showed AP firing rate's change during exposure to forskolin (10 μ M) and forskolin plus JWH133 (10 μ M). (d) Statistical analysis showed that forskolin increased AP firing rate, and in the presence of forskolin, JWH133 failed to reduce AP firing rate.

mode, JWH133 (10 μ M) still significantly reduced DA neuron AP firing rate (inhibitory rate was $11 \pm 2\%$, $n = 21$ from 6 mice, $p = 0.0001$, paired *t*-test, Fig. 10A), while without this pharmacological cocktail, JWH133-induced reduction in AP firing rate was $18 \pm 4\%$ ($n = 21$ from 8 mice, $p = 0.0002$, Fig. 10B). Although the block of synaptic transmission did reduce the inhibitory rate by JWH133 from $18 \pm 4\%$ (without synaptic block) to $11 \pm 2\%$ (with synaptic block), the difference of the inhibitory rate between these two groups ($18 \pm 4\%$ vs. $11 \pm 2\%$) was not statistical significant ($p = 0.165$, unpaired *t*-test). These results suggest that JWH133-induced mild reduction of presynaptic glutamate release seems not to play an important role in the mediation of JWH133-induced reduction in DA neuron firing. Second, we selectively blocked postsynaptic CB₂R-G protein function by addition of 600 μ M GDP- β -S, an intracellular G-protein blocker, into the recording electrode, thus allowing GDP- β -S to infuse into the recorded neuron after converted into whole-cell recording mode. As shown in Fig. 10, in the presence of intracellular GDP- β S, bath application of 10 μ M JWH133 failed to alter AP firing rate in VTA DA neurons (from 1.97 ± 0.39 to 2.03 ± 0.44 Hz, $p = 0.44$, paired *t*-test, $n = 7$ from 3 mice, Fig. 10C). However, in the absence of intracellular GDP- β S, bath application of 10 μ M JWH133 reduced VTA DA neuronal rate (from 1.99 ± 0.23 to 1.70 ± 0.22 Hz, $p = 0.004$, paired *t*-test, $n = 14$ from 4 mice, Fig. 10D). Statistical analysis showed that under whole-cell recording conditions (within 5 min after converted into whole-cell configuration), JWH133 failed to reduce AP firing rate in the presence of intracellular GDP- β S (Fig. 10E), while JWH133 reduced AP firing in the group neurons that did not contain intracellular GDP- β S (Fig. 10F). These results suggest that the intrinsic mechanism (CB₂R-mediated enhancement of M-current) predominantly underlies the JWH133-induced reduction of VTA DA neuron excitability.

4. Discussion

In this study, we find that acute exposure of the CB₂R agonist, JWH133, reduces DA neuronal excitability in the VTA, which is mainly mediated through an intrinsic mechanism. Although JWH133 mildly

reduces presynaptic glutamate release probability and results in a decreased glutamatergic synaptic transmission in VTA DA neurons, this may play little role in JWH133-induced reduction of VTA DA neuron firing because pharmacological block of synaptic transmission fails to prevent JWH133-induced inhibitory effect. JWH133 activates a postsynaptic CB₂R-G_{i/o} signaling cascade, reduces intracellular cAMP levels, in turn enhances M-type K⁺ channel function, and leading to the reduction of VTA DA neuronal excitability. Through this intrinsic mechanism, CB₂Rs significantly reduce VTA DA neuronal excitability, which may underlie CB₂R-mediated modulations in mesolimbic DA system function and DA-related behavior and diseases.

VTA DA neuronal firing is controlled by excitatory and inhibitory synaptic innervations. Modulations of these synaptic inputs alter neuronal excitability. Numerous G-protein-coupled receptors such as opioid, 5-HT₂, metabolic glutamate, GABA_B and CB₁ receptors modulate synaptic transmission by alteration of presynaptic neurotransmitter release. Emerging evidence demonstrates that activation of CB₂Rs modulates synaptic transmission in the entorhinal cortex [41], transfected hippocampal Dual neurons [42] and hippocampal pyramidal neurons from hippocampal slice cultures [43]. Thus, it is reasonable that CB₂Rs are expressed in presynaptic terminals/boutons and modulate neurotransmitter release. Indeed, in this study, we found a mild, but statistically significant, reduction in the frequency of mEPSC after exposure to JWH133 (10 μ M), suggesting that JWH133-induced reduction in VTA DA neuronal excitability may be mediated, at least in part, through a reduction of glutamatergic synaptic transmission onto DA neurons. However, additional experiments showed that pharmacological block of glutamatergic and GABAergic transmissions by perfusion of NBQX, APV and picrotoxin to VTA slice failed to prevent JWH133 (10 μ M)-induced reduction in DA neuron firing rate. These results suggest that JWH133-induced the mild reduction of presynaptic glutamate release does not contribute to the JWH133-induced reduction in DA neuron firing under our experimental conditions. Previous report has shown that the activation of CB₂Rs reduced inhibitory synaptic transmission in the entorhinal cortex [41], but this study did not identify the cellular location of the CB₂Rs involved [41]. Thus, the pharmacological effects

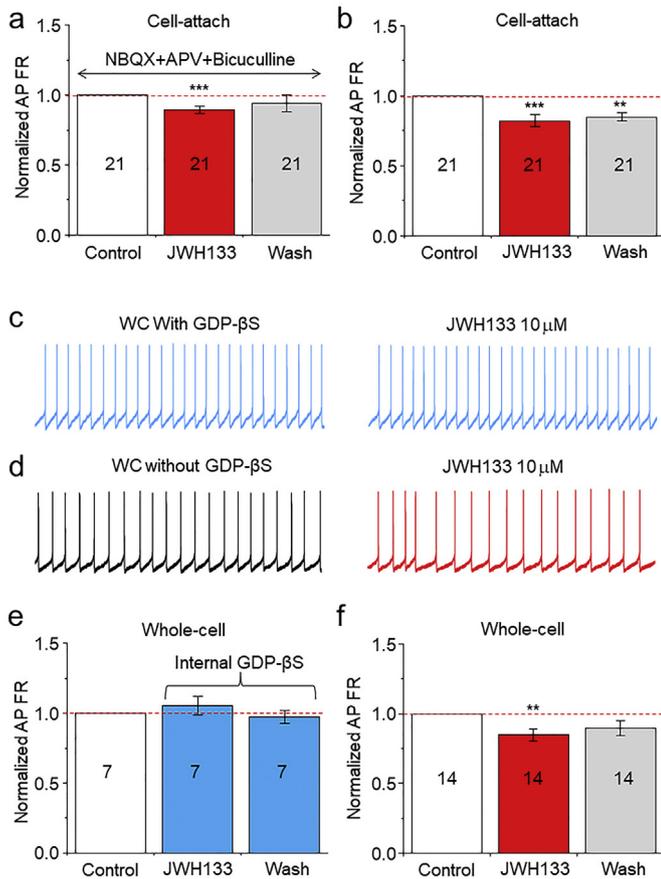


Fig. 10. Determination of the role of synaptic and intrinsic mechanisms in JWH133-induced reduction of VTA DA neuronal excitability. (a) Bar graph summarizes data that showed the alteration of AP firing rate before and after JWH133 exposure. The VTA DA neuron AP firing was recorded using cell-attach recording mode in the presence (a) and absence (b) of cocktail pharmacology (NBQX+APV + PTX) to block synaptic transmission. (c) Representative typical traces showing the AP firing measured by whole-cell recordings in current-clamp mode before (left) and after (right) JWH133 exposure. In these experiments, the GDP- β S (600 μ M) was added into recording electrode, and converted to whole-cell recording mode allowed GDP- β S infusion into recorded cells, and data showed that JWH133 failed to alter AP firing rate. (d) Representative typical traces in control group showing the AP firing measured by whole-cell recordings in current-clamp mode before (left) and after (right) JWH133 exposure without GDP- β S in the pipette solution and showed that JWH133 reduced AP firing rate. (e)/(f) Bar graph summarizes pooled data from (c) and (d) demonstrating that JWH133 significantly reduced AP firing rate in control (d), but not GDP- β S-treated neurons (c). ** $p < 0.01$, *** $p < 0.001$, compared to AP firing before JWH133 exposure.

reported may be caused by CB₂R expressed in either cortical neurons or microglia [44] since microglia are capable of modulating synaptic plasticity [45]. Another report demonstrated that selective activation of neuronal CB₂R reduced inhibitory synaptic transmission via a presynaptic mechanism in transfected hippocampal Duald neurons prepared from CB₂R^{-/-} mice [42]. Compared to these two reports, we found that CB₂R mildly reduced glutamatergic, but not GABAergic transmission in mouse VTA DA neurons. This discrepancy may be due to several reasons. First, different brain regions or neuronal phenotypes may have varied levels of CB₂R expression [46]. This is supported by our recent finding that CB₂R are highly expressed in the midbrain and moderately expressed in the prefrontal cortex (PFC), while the expression in the striatum is very low [25]. Second, brain CB₂R expression is inducible and dynamic. For example, repeated cocaine administration or self-administration significantly up-regulates brain CB₂R expression in the PFC, striatum and midbrain, as well as VTA neurons in rats [47]. Similarly, chronic, rather than acute, treatment of the slices with JWH133 for 7 days significantly enhanced excitatory synaptic transmission,

suggesting that chronic treatment of JWH133 may up-regulate CB₂R expression in the hippocampus [43]. Third, different CB₂R agonists, with varying pharmacological actions, were used in separate studies. These pharmacological variances, resulting in the activation of diverse intracellular signal pathways [48], may explain these inconsistent electrophysiological findings. Collectively, our data suggest that glutamatergic presynaptic terminal/boutons may express very low level of CB₂R, where they mildly modulate glutamate release probability, but contribute little to the alteration of DA neuron firing rate. Interestingly, we did not see a significant effect of JWH133 on mIPSCs, eEPSCs and eIPSCs in VTA DA neurons, which is consistent with previous report that activation of CB₂R modulated glutamatergic, but not GABAergic synaptic transmission in hippocampal slices [43]. The reason remains unknown, it may suggest that CB₂R are mainly distributed on the neuron somato, and only limited CB₂R are expressed on glutamatergic presynaptic terminals/boutons in VTA DA neurons. Our previous work supports this idea [25] as we showed predominant expression of CB₂R on the VTA DAergic cell body.

In addition to presynaptic effect, VTA DA neuronal excitability is also profoundly modulated by intrinsic postsynaptic mechanisms, including the linkage of G-protein-coupled receptors to ion channels located on the cell membrane. CB₂R are G-protein-coupled receptors that, when activated, alter G-protein (G_i/G_o)-coupled intracellular signaling cascades [49]. Emerging lines of evidence suggest that in VTA DA neurons, G-protein-coupled receptor signaling modulates neuronal excitability through several types of ion channels, e.g., G-protein-coupled inwardly rectifying K⁺ channels (GIRK) [50–52], M-type K⁺ channels [32,53,54], A-type of K⁺ channels [55], and L-type voltage-gated Ca²⁺ channels [56]. In the present study, we found that JWH133 reduced neuron firing in acutely-dissociated VTA DA neurons by prolonging AP initiation, enhancing AHP amplitude and hyperpolarizing membrane potential, suggesting a potentiation of M-type K⁺ current. Under voltage-clamp mode, we measured M-current in dissociated VTA DA neurons, and found a CB₂R-dependent enhancement of M-current amplitude by JWH133, confirming that CB₂R modulate M-current. M-channels are voltage-gated K⁺ channels that strongly activate on depolarization to repolarize the cell, and potentially regulate neuronal excitability [57,58]. Since M-type channels are activated at a membrane potential near the threshold for action potential initiation, it has a strong influence on neuronal excitability and responsiveness to synaptic inputs [59,60], and plays a critical role in terminating bursting activity in hippocampal CA1 neurons [61]. Importantly, under resting physiological conditions, M-channels are not fully open. Single channel analysis of both native [53,59] and recombinant [62,63] Kv7.2/Kv7.3 channels demonstrates that the maximal channel open probability is <1.0, suggesting that there is a considerable range to fully activate M-channels, and therefore, regulate neuronal activity. It was reported that VTA DA neurons express M-type K⁺ channels [32] and these channels play an important role in controlling VTA DA neuronal excitability [64]. It has been reported that activation of G-protein-coupled receptors increases M-currents in hippocampal neurons, by administration of somatostatin [65], corticostatin [66], or dynorphin [67]. This G-protein-mediated enhancement of M-current is important since it reduces neuronal excitability and counteracts convulsions. Indeed, it was also reported that the M-current enhancer, retigabine, shifts the current-voltage curve to the left (so that the channels open at more hyperpolarized membrane potentials) and increases the maximal opening probability [68,69], therefore, producing a strong suppression of neuronal firing [68] and exhibiting a potent anticonvulsant effect [70]. We also compared the effects of JWH133 and retigabine on M-current and VTA DA neuronal firing. We found that JWH133 and retigabine enhanced M-current amplitude and reduced neuronal firing rate. However, in the presence of M-current blocker (XE991), JWH133 failed to inhibit VTA DA neuron firing. These results suggest that CB₂R-mediated enhancement of M-current is an important intrinsic mechanism underlying the CB₂R-

mediated reduction in VTA DA neuronal excitability. Then, we further examined the signaling between CB₂R activation and M-current enhancement. Based on the previous studies, activation of CB₂Rs primarily initiates two signal pathways, MAPK/ERK and cAMP/PKA [37–39,48]. Considering cAMP plays an important role in modulation of M-current [40] and neuronal firing [71,72], we tested whether or not CB₂R-induced reduction of cAMP contributes to the enhanced effect of M-current. To reach this goal, we applied forskolin, a cAMP enhancer to the recorded cell to increase intracellular cAMP levels [73]. We tried to address 3 questions: [1] Does forskolin suppress M-current? [2] Does forskolin increase VTA DA neuron firing? [3] Does forskolin prevent JWH133-induced inhibition of VTA DA neuron firing? Our data clearly showed that forskolin (10 μM) reduced M-current and increased VTA DA neuron firing. In the presence of forskolin, JWH133 failed to potentiate M-current and reduce AP firing. These results suggest that CB₂R-G-protein-cAMP pathway likely plays a critical role in modulation of M-current and consequently neuronal excitability. Thus, under our experimental conditions (in vitro preparations), the activation of somatodendritic CB₂Rs on VTA DA neurons reduces intracellular cAMP levels and enhances M-current amplitude, leading to a reduction in neuronal firing and excitability. However, in vivo conditions may be more complex, for example CB₂Rs can modulate neuronal excitability by acting on glial cells [74,75], by modulating endogenous cannabinoid substances or even by acting on peripheral cells.

Our findings that the CB₂R-mediated reduction of VTA DA neuronal excitability, the underlying intracellular signaling (cAMP) and target (M-channels) have translational significance. Accumulating lines of evidence demonstrate that CB₂Rs mediate a variety of important modulations in DA-associated behaviors [76] including food intake, body weight [77–80], depression [81], anxiety [14,82], and schizophrenia-like behavior [15,83]. Recent reports emerging from several labs, including ours, have shown brain CB₂Rs play a pivotal role in the elimination of cocaine, alcohol and nicotine addiction [84–86]. These lines of evidence strongly suggest an important impact of CB₂Rs in the mesocorticolimbic system as well as critical roles in various brain functions including psychiatric, cognitive and neurobiological activity. The finding of the CB₂R-mediated reduction of midbrain DA neuronal excitability provides an insight into understanding of the roles played by CB₂Rs in the above mentioned DA-associated function and diseases. When compared to CB₁Rs, brain CB₂Rs exhibit highly translational potential to be used in clinical practices since the following unique features: [1] Lower expression levels, suggesting that these receptors may not mediate the effect of cannabis under normal physiological conditions; [2] Highly inducible, meaning that under some pathological conditions (e.g., addiction, inflammation, stroke, schizophrenia, stress, anxiety, etc.), CB₂R expression is enhanced in the brain [87], suggesting a close relationship between the alteration of CB₂R expression/function and various psychiatric and neurological diseases; and [3] Exhibit special distribution, given that CB₂Rs are chiefly expressed in neuronal somatodendritic areas [8] (postsynaptic) but CB₁Rs are predominantly expressed on neuronal terminals, especially on GABAergic terminals (presynaptic), which leads to some opposing effects after activation by these two receptor subtypes [88]. Considering these characteristic features, the mesolimbic CB₂R appears to be an important substrate for neuroprotection [89], and targeting CB₂Rs or manipulations of their signaling (cAMP/PKA) and target (M-channels) will likely offer a novel therapeutic strategy for treating neuropsychiatric and neurological diseases without typical CB₁R-mediated side-effects.

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ebiom.2019.03.040>.

Conflicts of interest

All authors have nothing to disclose.

Author contributions

Z.M., F.G., B.L., M.G., Z.L., D.C., X.M., S.Q., Y.Z., Z.X. conducted the experiments, analyzed data, prepared original figures, and revised the manuscript. Z.X. and J.X. participated in the experimental designs, analyzed the data, and revised the manuscript. J.W. designed and conducted the experiments, analyzed data, finalized figures, and wrote the manuscript.

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