

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23

**Time to branch out? Application of hierarchical survival models in plant phenology**

Sarah C. Elmendorf <sup>1,2,\*</sup>, Theresa M. Crimmins <sup>3,4</sup>, Katharine L. Gerst <sup>3,4</sup>, and Jake F. Weltzin <sup>3,5</sup>

<sup>1</sup> Institute for Arctic and Alpine Research (INSTAAR), University of Colorado, Boulder, CO, USA

<sup>2</sup> Ecology and Evolutionary Biology Department, University of Colorado, Boulder, CO, USA

<sup>3</sup> National Coordinating Office, USA National Phenology Network, Tucson, AZ, USA

<sup>4</sup> School of Natural Resources and the Environment, University of Arizona, Tucson, AZ, USA

<sup>5</sup> U.S. Geological Survey, Ft. Collins, CO, USA

\*Corresponding author: Sarah Elmendorf ([sarah.elmendorf@colorado.edu](mailto:sarah.elmendorf@colorado.edu))

This draft manuscript is distributed solely for purposes of scientific peer review. Its content is deliberative and pre-decisional, so it must not be disclosed or released by reviewers. Because the manuscript has not yet been approved for publication by the U.S. Geological Survey (USGS), it does not represent any official USGS finding or policy.

**DECLARATIONS OF INTEREST:** none

24 **ABSTRACT**

25

26 The sensitivity of phenology to environmental drivers can vary across geography and species.  
27 As such, models developed to predict phenology are typically site- or taxon-specific. Generation  
28 of site- and taxon-specific models is limited by the intensive in-situ phenological monitoring  
29 effort required to generate sufficient data to parameterize each model. Where in-situ  
30 phenological observations exist, the data are often subject to analytical issues due to the limited  
31 duration of any individual monitoring program, spotty site- and species- level coverage, lack of  
32 standardized methodology, and infrequent or variable census intervals. Together, these  
33 characteristics constrain our ability to make phenological inferences outside of select sites and  
34 taxa where long-duration, intensive monitoring has occurred.

35

36 In this study, we leveraged two national, standardized phenology datasets to develop a multi-  
37 species and multi-site state-space survival model of the onset of deciduous tree and shrub  
38 spring (leaf out) and fall (leaf-color) events across temperate ecoregions of the United States.  
39 We used data from two national-scale phenological databases, a 9-year, broadly distributed  
40 dataset from the USA National Phenology Network and a 4-year dataset from the National  
41 Ecological Observatory Network, to quantify regional and interspecific variation in sensitivity to  
42 environmental drivers for both spring and fall leaf phenophases. Spring leaf out was generally  
43 promoted by longer days, spring growing degree day accumulation, overwinter chilling, and was  
44 suppressed by frost events, whereas fall leaf color was promoted by shorter days and cold  
45 accumulation. The sensitivity to most environmental drivers tended to be more variable among  
46 species than among the regions as defined here (EPA ecoregions of North America, excluding  
47 desert and tropical areas). The results of this study lay the groundwork for incorporating the  
48 growing collection of phenological observations into a generalized framework for predicting the  
49 transition states for any species, in any location.

50

51

52 **KEYWORDS:** citizen science, phenology, temperature, day length, USA-NPN, NEON

53

54 **HIGHLIGHTS:**

- 55 • Hierarchical survival models of plant phenology were developed
- 56 • Citizen science and national-scale monitoring phenology data were analyzed
- 57 • Chilling, spring warming, longer days, and less frost promote spring leaf out
- 58 • Cold accumulation and shorter days promote fall leaf coloration.

59

## 60 **1. INTRODUCTION**

61

62 The ability to predict the timing of life cycle events for individual species at given locations is  
63 beneficial to many aspects of resource and species management (e.g. optimally timing of land  
64 prescribed burns or chemical treatments; (Enquist et al., 2014)) and human health (e.g.  
65 forecasting the onset or peak of airborne pollen in allergenic taxa (Siljamo et al., 2013).

66 Understanding the drivers to phenology across species also improves our ability to anticipate  
67 future changes to ecosystem services as climate conditions change, due the impact of plant  
68 phenology on carbon dynamics (Richardson et al., 2013), hydrology (Morissette et al., 2009), and  
69 albedo (Williamson et al., 2016).

70

71 In recent years, many predictive models have been developed to elucidate the drivers of plant  
72 phenology, ranging from empirical approaches such as regression (Hufft et al., 2018) to more  
73 complex process-based models that attempt to explain underlying physiological responses  
74 (Basler, 2016). Model construction is frequently data-limited; phenological datasets with both

75 broad taxonomic and geographic coverage that are consistently sampled over decades do not  
76 exist. Accordingly, phenology modeling efforts typically fall into one of a few categories: those  
77 that analyze one or more species at a single site or small region (Bertelsen's data from the  
78 Santa Catalina mountains; the Marshall phenology record from central England, or Thoreau's  
79 Concord records (Crimmins et al., 2010; Miller-Rushing and Primack, 2008; Roberts et al.  
80 2015), those that combine phenological observations across different species, regions, or  
81 sampling to describe general patterns using meta-analysis type approaches (e.g Cook et al.,  
82 2012; Oberbauer et al., 2013), and those that use space-for-time substitution to parameterize  
83 models for single species using a relatively short time-series across multiple sites (Liang and  
84 Schwartz, 2014; Melaas et al., 2016; Taylor et al., 2019). Relatively simple phenology models  
85 often show equivalent predictive performance with more complex models. However, with all  
86 models, transferability of fitted parameters among sites is imperfect (Basler, 2016).

87

88 An additional challenge in modeling phenology is that observations often are conducted  
89 infrequently because of the effort required in regular censusing, introducing uncertainty into the  
90 timing of when an event occurred. Because of this, phenological models have typically been  
91 developed separately for the few species or handful of sites with a rich phenological record,  
92 which excludes the remaining "long-tail" of phenological observations: those collected over a  
93 short duration; those that have few replicates for any species or location; and those that have  
94 lower sampling frequency. For phenological studies, we are fortunate that many of these "long-  
95 tail" datasets have been collated and publicly archived, making such observations readily  
96 available for inclusion in analyses. What remains to be developed is an analytical approach that  
97 leverages these data, which can broaden the diversity of species, sites, and time-frames  
98 represented, and can enable national-scale inference.

99

100 Survival analysis approaches are a promising solution for capitalizing on the long tail of unused  
101 phenological data. Survival models are a class of statistical models that are well-suited to  
102 accommodating the typical shortcomings of phenology datasets and that have the potential to  
103 outperform traditional regression models in predicting phenological events (Templ et al., 2017).  
104 Survival analysis has a long history in engineering studies of failure time, as well as in the  
105 medical literature in estimating time to death or disease recurrence where observations are  
106 right-censored (e.g. Cox 1972). The analogy to plant monitoring data is clear: Like patients,  
107 individual plants may be monitored at long or irregular intervals and in some cases the event of  
108 interest occurs after the monitoring period is over. Only a handful of studies have used survival  
109 approaches to model climate drivers of plant phenology, despite its potential (e.g. Allen et al.,  
110 2015; Diez et al., 2014; Templ et al., 2017, Terres et al., 2013).

111  
112 We additionally considered the best ways of including data from multiple species and multiple  
113 sites. Interspecific variation in phenology dates can be substantial, commonly on the order of  
114 several weeks (Lechowicz, 1984). Further, individual species can show different sensitivity to  
115 environmental drivers across locations (Ma et al., 2018; Parker et al., 2017; Prev y et al., 2017;  
116 Way and Montgomery, 2015), but see MacKenzie et al. (2018) for a counterexample. However,  
117 despite clear differences among species in the dates of phenological transitions, experimental  
118 work has shown the phenology of a diversity of deciduous taxa responds similarly to a common  
119 set of environmental cues (Flynn and Wolkovich, 2018) and many successful models have been  
120 developed by aggregating phenological data using space-for-time substitution. The potential for  
121 heterogeneity in sensitivity to environmental drivers among species and regions argues for  
122 phenological forcing models that specifically allow for this variability. At the other extreme,  
123 modeling each species and region entirely separately dramatically reduces sample sizes.  
124 Therefore, to the degree which cues are common across species and regions, these common  
125 characteristics should be leveraged to improve models for all species and regions.

126

127 Using hierarchical models, it is possible to incorporate variation among species and regions as  
128 random effects (Clark 2007, Bolker 2008). In this framework, the estimated sensitivity of any  
129 individual species or region is influenced by the sensitivities of other taxa and locations,  
130 because inferences on species- and region-specific coefficients “borrow strength” from species  
131 and regions. Using a multi-species, multi-region approach, the modeled sensitivity of any  
132 individual species or region reflects both the sensitivity that would be estimated analyzing each  
133 species or region separately and the average sensitivity over all species and regions (Gelman  
134 and Hill, 2007). Thus, integrating survival models into a hierarchical framework further broadens  
135 the ability to incorporate additional datasets into the analysis by accommodating observations  
136 made on different species and regions.

137

138 New, geographically extensive and taxonomically diverse in-situ phenological datasets collected  
139 over broad geographic regions broadens possibilities for developing and improving phenological  
140 models. The USA National Phenology Network (USA-NPN) and the National Ecological  
141 Observatory Network (NEON) are national-scale programs with a primary (USA-NPN) or  
142 substantial (NEON) focus on collecting phenological data using concordant, standardized  
143 protocols that have substantially increased the availability of in-situ phenological observations  
144 across the United States over the past decade (Denny et al., 2014; Elmendorf et al., 2016). The  
145 USA-NPN dataset has been used for numerous applications to date, including the identification  
146 of climate drivers of phenological transitions for individual species (e.g. Piao et al. 2015, Mazer  
147 et al. 2015, Park and Mazer 2018; Melaas et al., 2016; Taylor et al. 2019), understanding spatial  
148 and temporal phenological trends (e.g. Pearse et al. 2017, Liang 2019), and evaluation of  
149 remotely sensed data (e.g. Peng et al., 2017; Zhang et al., 2018). However, none to date have  
150 leveraged either the USA-NPN nor the NEON datasets to create hierarchical multi-species  
151 models across broad geographic regions.

152

153 In this study, we outline a novel analytical approach for analyzing this growing collection of  
154 phenology observations across a broad array of ecosystems and species. The objective of our  
155 work was to develop hierarchical multi-region, multi-species survival models of the onset of  
156 spring (leaf out) and fall (leaf coloration) for over 100 plant species within 15 temperature  
157 ecoregions regions of the U.S. The resulting models demonstrate common drivers of deciduous  
158 spring and fall phenology across species in temperate regions of the U.S. and provide estimates  
159 of sensitivity for each species and region.

160

## 161 **2. MATERIALS and METHODS**

162

### 163 **2.1 Model overview**

164 Survival analysis is a modeling approach robust to the challenges that phenology data pose.  
165 First, phenology data are essentially time-to-event data, and therefore show different statistical  
166 properties than other typical ecological parameters such as size, counts, or concentration.  
167 Second, the drivers of phenological transitions vary temporally so that including “average”  
168 weather (e.g., April mean daily temperature) as a phenological predictor misrepresents the  
169 biology of the system. For example, recent weather events are likely to have an outsized  
170 influence on the near-term rate of phenological progression, and weather after a phenophase  
171 transition obviously cannot have any effect on the date of the (past) event. Third, in the absence  
172 of continuous monitoring, the date on which phenological transitions occur is typically reported  
173 with varying degrees of uncertainty, depending on the frequency of observation. Phenological  
174 datasets – especially those collected opportunistically or infrequently by volunteers -- can suffer  
175 from both relatively long periods with no observations as well as variability in temporal  
176 frequency of observations. Survival analysis, which can accommodate temporally varying

177 forcing variables and any intercensus interval, is well-suited to data with these characteristics  
178 (Templ. et al., 2017).

179

180 Here, we implement survival modeling using discrete time models where the time-step is a  
181 single day, which is typically the smallest intercensus sampling interval for plant phenology.

182 Borrowing from the nest-survival literature (Royle and Dorazio, 2008), we categorize a single

183 plant's spring (or fall) phenology as a state change from one phase (e.g., without leaves) to

184 another (e.g., with leaves). For the purpose of model development, we classify the states as a

185 binary variable that starts in state '1' and has a modeled probability of transitioning to state '0' on

186 each day of year. Mathematically, we can consider this daily probability of changing state as a

187 daily 'hazard' rate, or the complement of the daily 'survival' rate  $\varphi_d$ , which is the probability of

188 staying in the same (pre-transition) phenophase state on any date.

189

190 Thus, on each day ( $d$ ), for each individual ( $i$ ),

191 *Eq (1):*

$$192 \quad P(\text{State}_{i,d} = 1 | \text{State}_{i,d-1} = 1) = \varphi_{id}$$

193

194 Once transitioned, the probability of remaining in the terminal state is 100%, i.e.

195 *Eq (2):*

$$196 \quad P(\text{State}_{i,d} = 0 | \text{State}_{i,d-1} = 0) = 1.$$

197

198 After transitioning to the (seasonally) terminal state ('0'), reversals are not possible in this

199 framework (i.e., this is a single-season model). The influence of factors that vary at the level of

200 an individual (e.g. geolocation, species, or traits) and/or time period within individual (e.g. daily

201 weather at an individual's location) on phenological timing can be modeled by adding these  
202 factors as predictors of the daily survival rates.

203

204 We combined data across multiple taxa and regions into a single hierarchical, multi-species,  
205 multi-region space-state model separately for spring and for fall phenological events. We used a  
206 state-space modeling framework where the daily probability of transitioning from the initial state  
207 (leafless for the spring model; lacking colored leaves for the fall model) to the terminal state  
208 (with leaves for the spring model; with colored leaves for the fall model) is a function of  
209 environmental drivers that occurred in the time prior to each event. The sensitivities of each  
210 species and region to each environmental driver were modeled as random effects. Further  
211 details on the statistical modeling approach are provided in Appendix A.

212

## 213 **2.2 Environmental drivers**

214 To assemble a suite of candidate environmental drivers of spring leaf out and fall leaf color to  
215 incorporate into multispecies, multi-region models, we surveyed the literature to prioritize the  
216 putative drivers (Appendix B). Based on that analysis, we selected growing degree days (GDD),  
217 daylength, chilling, precipitation and frost as environmental forcing variables for the spring leaf  
218 model. Environmental forcing variables for the fall leaf color model included cold degree days  
219 (CDD), precipitation and daylength. Climate data used to derive these environmental forcing  
220 variables were extracted from Daymet (Thornton et al., 2018).

221 Environmental drivers used as covariates were calculated using an accumulation window  
222 approach, reflecting cumulative and dynamic effects on organismal biology. Accumulation  
223 windows can be calculated over a moving window period (e.g. the sum of heat over an n-day  
224 moving window) or with a fixed start date (e.g. the sum of heat since Jan 1). While the latter is  
225 more common, the calendar date selected (usually Jan 1) is fairly arbitrary and a wide range of

226 parameterizations have been used successfully. After considering both the geographic spread  
227 of our data and parameterizations used in previous studies, we included as environmental  
228 drivers the sum of frost events, cumulative precipitation, and cold degree days (CDD, base 20)  
229 over a moving window 14 days to one day prior to each modeled timepoint, the sum of chilling  
230 hours using the Utah model (Richardson et al., 1974), over a moving window 135 days to 45  
231 days prior to each modeled timepoint, and growing degree days (GDD, base 0) over a moving  
232 window 45 days to one day prior to each modeled timepoint and (Diez et al., 2014; Gill et al.,  
233 2015; Gunderson et al., 2012), and the prior day's daylength (Appendix C). These windows  
234 generally created forcing variables for growing degree days, cold degree days, frost, and chill  
235 accumulation that were changing unidirectionally over the spring and fall periods. All covariates  
236 were scaled to mean 0, unit variance prior to their inclusion in the models to facilitate model  
237 interpretation and convergence.

238

### 239 **2.3 Filtering and pre-processing of phenological data**

240 We used publicly available datasets from the USA-NPN (USA National Phenology Network, 2018)  
241 and NEON (National Ecological Observatory Network, 2018) to predict the onset of leaf out in  
242 spring and leaf coloration in fall for deciduous tree and shrub taxa over the temperate regions of  
243 the US. The USA-NPN developed standardized monitoring methods for plants and animal taxa  
244 that include the assessment of the phenological status (presence or absence) of carefully  
245 described, recognizable phenophases (Denny et al. 2014). Citizen science monitoring launched  
246 in 2009 and is conducted by tens of thousands of professional scientists and volunteers. NEON  
247 is a national-scale ecological observatory which conducts standardized measurements of  
248 multiple ecological parameters in 47 terrestrial and 34 aquatic sites across the US. The  
249 program has just entered its operational phase, but plant phenology measurements at some  
250 sites are available from as early as 2014. USA-NPN and NEON monitoring protocols use  
251 identical phenophase definitions and are status-based, such that the raw data on each day of

252 year is an observation of whether and individual is in a particular phenophase (i.e., has leaves)  
253 or not (has no leaves) (Denny et al. 2014). At each NEON terrestrial site, plant phenology is at  
254 regular intervals by trained technicians. We accessed all available data using the USA-NPN and  
255 NEON APIs (application programming interface) and retained observations for species denoted  
256 as “Deciduous Trees and Shrubs” by the USA-NPN.

257  
258 Response variables in the space-state model include, for each monitored individual, in each  
259 year, the latest day of year in which an individual was observed to be in the initial state (i.e.,  
260 leafless for the spring leaf model; having green leaves for the fall color model) and (optionally)  
261 the earliest day of year in which an individual was observed to be in the terminal state (with the  
262 phenophase “leaves” for the spring leaf model, and having the phenophase “colored leaves” for  
263 the fall color model). We converted the raw status observations into 'last observed in initial state  
264 (last no)' values and 'first observed in terminal state (first yes)' values based on the annual time-  
265 series (Appendix D). In some cases, absence of the phenophase was not recorded (i.e., no 'last  
266 no' values could be extracted from the observations). For these individuals, we assumed the  
267 individuals lacked leaves in the middle of winter (Day of Year [DOY]=1), and had no colored  
268 leaves in the middle of summer (DOY = 180). Because the survival analysis approach we took  
269 can accommodate any inter-census interval, we did not further filter observations to those with a  
270 particular duration between the 'last observed no' and 'first observed yes' values, as is typically  
271 necessary for working with phenology data that include some observations with large inter-  
272 census intervals.

273  
274 We assigned each observation based on its location to an EPA level II ecoregion (United States  
275 Environmental Protection Agency, 2018, discarding observations that fell into non-temperate  
276 (i.e., Mediterranean, tropical and desert-type) regions. In total, we used 16,942 observations  
277 (individual\*year combinations) representing 128 species in the spring model and 16,244

278 observations representing 112 species in the fall model (the full list of species included can be  
279 found in supplemental tables S1 and S3). Both models covered records across 15 ecoregions of  
280 the US (the full list of regions used can be found in supplemental tables S2 and S4) with data  
281 from 2009-2017 (USA-NPN) or 2014-2017 (NEON).

282

## 283 **2.4 Model evaluation**

284

285 Because our data included uncertainty in the response variable (the true date of the  
286 phenophase transition may have occurred at any point between the observed last no and  
287 observed first yes), we could not rely on traditional methods such as RMSE to assess model fit.  
288 Instead, we include a visual assessment of model fit, plotting the model-predicted median  
289 phenophase transition DOY vs. observed date range ('last observed no' → 'first observed yes')  
290 for each individual\*year combination. We also calculated the ranked probability score (RPS)  
291 (Epstein, 1969; Murphy, 1970) for a quantitative assessment of relative model fit, following as  
292 similar example in Terres et al. (2013). Specifically, we calculated a relative ranked probability  
293 score ( $RRPS_i$ ) for each individual observation, defined as the average of the squared difference  
294 of the probability of having transitioned by any day of year ( $P_{i,t}$ ) and the actual state (0,1) over  
295 all dates prior to and including the 'last observed no' and after and including the 'last observed  
296 yes.'

297

$$298 \quad RRPS_i = \frac{(\sum_{t=minStart}^{t=lastNo} [P_{i,t}]^2 + \sum_{t=firstYes_i}^{t=maxEnd-1} [1 - P_{i,t}]^2)}{maxEnd - minStart - (firstYes_i - lastNo_i)}$$

299

300

301

302 where  $P_{it}$  is the modeled probability of having transitioned (state=1) by any given day of year (t).  
303 Because the state is an ordered variable and  $P_{it}$  is a monotonically increasing function,  $RRPS_i$   
304 scores are always higher for individuals with greater measurement uncertainty (i.e., more days  
305 between the 'last observed no' and 'first observed yes'). We averaged all  $RRPS$  measurements  
306 over all observations included in each model for an overall assessment of model quality ( $\overline{RRPS}$ ).  
307  $\overline{RRPS}$  scores are only valid when compared to the same input dataset, so, for example, a  
308 comparison of the  $\overline{RRPS}$  for the spring vs. fall models is not meaningful because the underlying  
309 measurement uncertainty of observations included in the two datasets differs. When applied to  
310 different models of the same dataset, lower  $\overline{RRPS}$  values indicate better model performance.

311

### 312 **3. RESULTS**

313

#### 314 **3.1 Model assessment**

315

316 For the spring leaf out model, we were able to fit a single model including growing degree days  
317 (GDD), daylength, chilling, precipitation and frost as environmental forcing variables as well as  
318 an intercept (baseline daily probability of remaining in the initial [leafless] state for the spring leaf  
319 model ( $\hat{R}$  for all parameters <1.1). Inclusion of frost days in the fall model created convergence  
320 problems. We also experienced a lack of convergence when we included both intercept and  
321 daylength as predictors in the same fall model. These issues with convergence were likely due  
322 to the large number of days prior to the phenological event that included 0 sum frost days. We  
323 therefore fit the fall model without frost as well as alternative forms of the fall model, including  
324 either an intercept or daylength but not both. Also, to parallel the fall model, we constructed  
325 alternative forms of the spring model with and without an intercept. Spring models that included  
326 an intercept showed substantially better fit ( $\overline{RRPS} = 0.026$ ) than those without ( $\overline{RRPS} = 0.112$ ).

327 The fall model that included daylength had a slightly better fit ( $\overline{RRPS}=0.063$ ) than the model that  
328 included an intercept ( $\overline{RRPS}=0.071$ ).

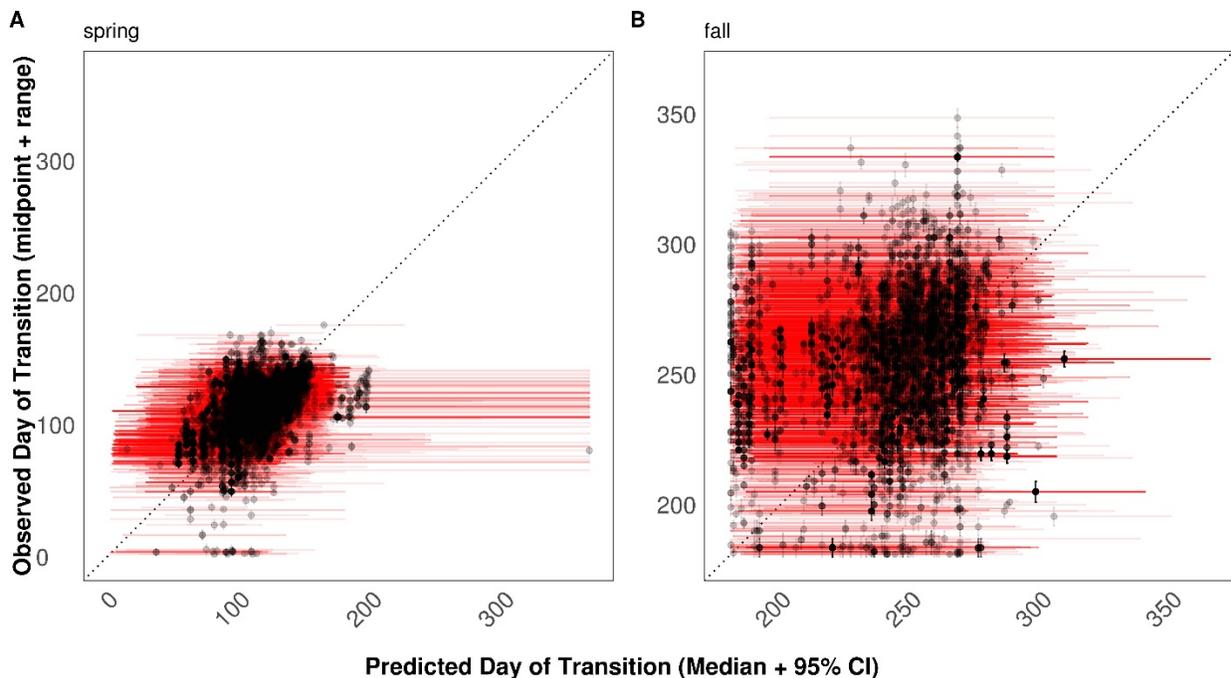
329

330 Visual assessment of both spring and fall models indicated better predictive performance of the  
331 spring vs. fall model (Fig 1). In particular, the fall model erroneously predicted a suite of fall  
332 coloration events to occur very early in the season (before DOY 200). Spring data showed  
333 better agreement between predicted and observed dates.

334

335

336



337

338

**Figure 1.** Predicted versus observed dates of spring leaf out for 128 species (A) and fall leaf coloration for 112 species (B). Figure shows median estimated date of leaf out or fall coloration (x axis, and horizontal 95% CI in red) against observed dates of transition (vertical black bars, lower = 'last observed no,' upper = 'first observed yes,' dot = midpoint). Perfect predictions would fall along the 1:1 line. For purposes of the figure, we included only observations with <10d intercensus intervals between the 'last observed no' and 'first observed yes,' though the model included all data.

339

340

341

### 342 3.2 Sensitivity to Environmental Drivers

343

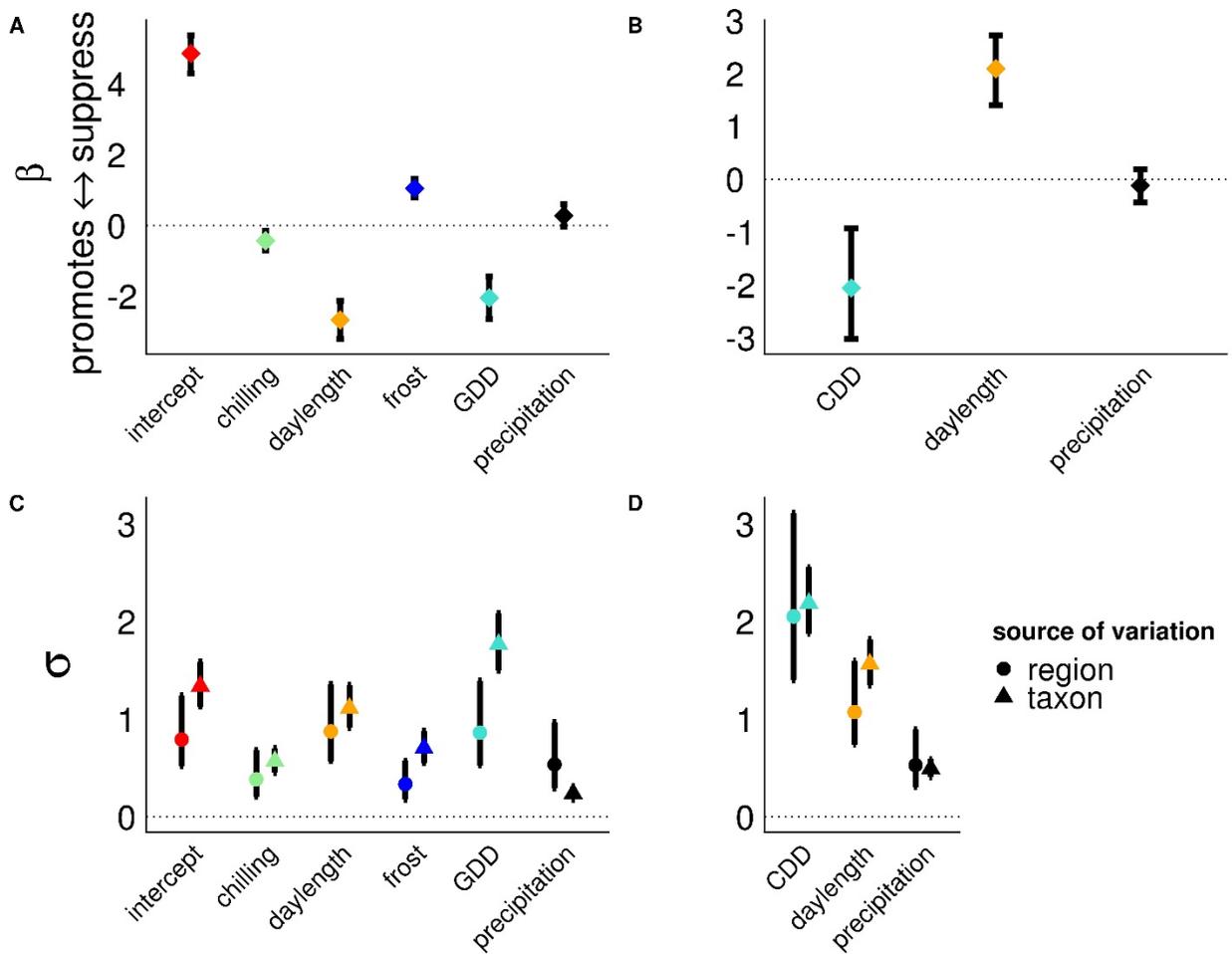
#### 344 3.2.1 Spring leaf out

345 On average, spring leaf out was promoted by longer days, higher growing degree day sums,

346 higher amounts of chilling, and was suppressed by recent frost events (Fig 2a). Precipitation

347 had a slight but nonsignificant suppressing effect on spring leaf out.

348



349

350

351

352

353

**Figure 2.** Mean sensitivities and variance in sensitivities among species and regions of deciduous trees and shrubs phenology to environmental drivers. Panels A (spring leaf out) and B (fall leaf coloration) show estimated mean sensitivity + 95% CI for each environmental driver, plus the baseline hazard rate (intercept). Positive coefficients indicate climate forcing variables that prolong the initial state (i.e., leafless state for spring; green-leaved state for fall), and therefore delay the onset of the phenological transition. Panels C and D show the variability in sensitivity of drivers for spring (C) and fall (D) models among geographic regions (squares) and taxa (triangles).

354 -

355

356 Models successfully generated estimates of sensitivity of each taxon and each region to the  
357 environmental drivers. With the exception of precipitation, sensitivities of leaf out to  
358 environmental drivers varied more among taxa than among regions (Fig 2c). To illustrate the  
359 ability of the models to generate estimates for the large diversity of taxa included here,  
360 sensitivities of spring leaf out to one environmental driver (GDD) is presented in Figure 3. The  
361 full suite of parameter estimates for all environmental drivers for each species and region for  
362 both the spring models are included in the Supplementary materials (Tables S.1-S.4).



**Figure 3:** Sensitivity of spring leaf out to GDD for each taxa (point estimate + 95% CI of each  $\beta_{GDD,species}$ ) term. Negative parameter estimates reflect species where GDD accelerates spring leaf out.

364

### 365 **3.2.2 Fall leaf coloration**

366

367 On average, fall leaf coloration was promoted by short days and higher cold degree day sums.  
368 Precipitation had a slight but nonsignificant suppressing effect on fall coloration (Fig 2B). For fall  
369 coloration, we again saw higher variability in sensitivity among species than regions to  
370 daylength, but more similar amounts of variability among species and regions for precipitation  
371 and cold sensitivity (Fig 2D). The full suite of parameter estimates for all environmental drivers  
372 for each species and region for both the spring models are included in Appendix F.

373

## 374 **4. DISCUSSION**

375

376 Recent growth and standardization of phenology observations across broad spatial scales over  
377 multiple years has the potential to enhance our understanding of the drivers of temperature  
378 plant phenology. Yet the patchiness of available data results in analytical challenges with  
379 respect to spatial, temporal, and taxonomic coverage. We applied a novel analytical framework  
380 to address these challenges and better understand the drivers of temperate leaf phenology and  
381 quantify sources of variability. On average, spring leaf out was accelerated by longer days,  
382 spring growing degree day accumulation, overwinter chilling, and was suppressed by frost  
383 events, whereas fall leaf color was accelerated by shorter days and cold accumulation.  
384 Comparisons across species and regions indicated climate sensitivity tended to be more  
385 variable among species than among regions.

386

#### 387 **4.1 Advantages of a hierarchical survival modeling approach**

388

389 We used hierarchical survival models to predict spring and fall phenological events across many  
390 taxa and regions. Space-state phenology models have been developed using datasets with  
391 frequent and regular census intervals (Allen et al., 2014; Diez et al., 2014; Xie et al., 2015).

392 These models can also be easily applied to datasets with variable and irregular observation  
393 frequencies. Such flexibility makes state-space approaches particularly well-suited to the data  
394 collected by volunteers, where infrequent and unevenly spaced and sampling intervals are the  
395 norm. This approach overcomes a common obstacle to embarking on a phenological campaign,  
396 namely, a requirement of high-frequency sampling regimes (Fitchett et al., 2015; Menzel, 2002).

397

398 In addition to the ability to make use of data with infrequent and/or variable intercensus-  
399 intervals, our approach has several advantages over common methods to analyze phenological  
400 data in terms of representing our understanding of the biological drivers of phenology. Modeling  
401 the daily probability of an event occurring permits us to incorporate weather up to but not  
402 beyond the date of a phenological transition as drivers of phenology, which is consistent with  
403 our understanding of the biological basis of phenological cues. Using a moving window for  
404 forcing accumulation also allows daily probabilities to vary non-monotonically over the growing  
405 season; for example, a late spring frost event could suppress the leaf out probability after that  
406 event occurs to levels below that before the frost event. Daily models require substantial data  
407 volumes to fit; models for only a few species would achieve convergence if each species were  
408 modeled separately. At the other extreme, combining datasets across taxa and regions (such as  
409 is commonly done in meta-analyses) ignores potential important variability in the sensitivities to  
410 environmental drivers among species and regions. Using a hierarchical framework allows us to  
411 leverage a large volume of data collected over many sites and species, despite the fact that the

412 dataset as a whole contains relatively few long time series of a single species at a given site.  
413 This approach allows us to gain holistic insight into general drivers of deciduous spring and fall  
414 phenology across temperate regions of North America, quantify the degree of variability among  
415 species and regions, as well as generate estimated phenological sensitivities for over 100  
416 species and 15 ecoregions.

417

#### 418 **4.2 Drivers of spring and fall phenology**

419

420 The dominant controls on deciduous tree phenology over most non-desert, temperate regions of  
421 the world are generally considered winter chilling, day length and temperature (Korner and  
422 Basler, 2010). Our analysis of over 100 deciduous taxa confirmed the importance of these  
423 drivers, showing that on average spring leaf out was stimulated by longer days, growing degree  
424 sums, and chilling. These results concur with those of Templ. et al. 2017, who found a strong  
425 positive effect of GDD on flowering using survival modeling, but a weak to insignificant effect of  
426 precipitation. The relationship between spring frost or precipitation and spring leaf out events  
427 has been less well-studied than that of temperature and daylength. Our results indicated a  
428 previously unappreciated role of spring frost in delaying leaf out and a slight trend (different from  
429 0 for a few taxa) of spring precipitation to delay leaf out. The magnitude of the negative  
430 (suppressive) effect of frost events was stronger than the positive (promoting) effect of  
431 overwinter chilling. The physiological (and ensuing demographic effects) of frost damage are  
432 well-documented (Inouye, 2000), so it is unsurprising that plants use recent frost events as a  
433 proximal cue for the emergence of sensitive tissues. Our leaf out results parallel a recent  
434 synthesis of flowering sensitivity to frost (Park and Mazer, 2018), with both studies highlighting  
435 the importance of frost as a driver of the timing of spring phenological events.

436

437 The direction of the mean sensitivities of fall leaf coloration to climate drivers was partially  
438 consistent with previous studies. Xie et al (2018) found rainfall in autumn cause earlier peak  
439 color, while we found a non-significant trend in the same direction. Previous research on the  
440 effects of temperature on the timing of fall onset have been conflicting, with evidence to support  
441 both advances and delays in leaf coloration with warmer temperatures under natural conditions  
442 (see Archetti et al., 2013 and references therein), whereas experimental manipulations tend to  
443 suggest a more consistent delaying effect of summer and fall temperature on senescence (Fu et  
444 al., 2018; Gunderson et al., 2012). The sign and strength of the relationship between  
445 temperature change and onset of fall can vary depending on the time of year and fall  
446 phenophase under consideration (Archetti et al., 2013)

447

448 We did not detect a strong role of precipitation in the onset of either spring or fall phenological  
449 events in this study. In temperate regions such as our study area, plants likely experience  
450 moisture stress or abundance through soil moisture, which is a function of soil properties and  
451 land cover as well as incoming precipitation (Pan et al., 2003). For example, Terrestrial  
452 biosphere models such as ORCHIDEE, CLM, LPJ-GUESS and Sheffield- DGVM commonly  
453 assume soil moisture physiologically constrains leaf phenology (Abramoff and Finzi, 2015).  
454 Improved satellite-based soil moisture products (e.g. Dorigo et al. 2017 or Reichle et al 2018),  
455 when generated on fine-spatial scales for a sufficient number of years, might provide a  
456 detectable soil moisture signal beyond the limited trends with precipitation found here.

457

458 Species showed consistently strong sensitivity to day length in both the spring (Tables S.1) and  
459 fall models (Table S.2). These correlative results are consistent with experiments that show  
460 photoperiod sensitivity to be fairly ubiquitous in spring events (Flynn and Wolkovich (2018). The  
461 relationship of daylength to fall coloration has not been as thoroughly studied experimentally;

462 however, our results concur with the hypothesis that day length is likely a primary regulator of  
463 fall phenology (Way and Montgomery, 2015).

464

#### 465 **4.3 Variability in sensitivity**

466

467 Our results demonstrate that sensitivity to environmental drivers varies both by species and by  
468 region. The inter-specific and inter-regional variation in sensitivity to environmental drivers found  
469 in this study helps explain the heterogeneity of results of previous studies of phenological  
470 sensitivity conducted on different species, which has limited transferability of phenological  
471 models parameterized at one location to other locations (Basler 2016). Previous studies have  
472 noted that interspecific variability can be as important as climate in predicting landscape level  
473 phenological patterns (Xie et al., 2015). Our results support an important role for species  
474 composition and location in understanding larger-scale phenological responses. Our process for  
475 selecting regions and species for inclusion must be kept in mind when interpreting the  
476 magnitude of the region and species variability. We constrained our geographic regions to  
477 exclude desert and tropical ecoregions and included species only within a single life form. It is  
478 likely that broadening the species pool or geographic locations included would increase the  
479 amount of interspecific or inter-regional variability detected. Unsurprisingly, parameter  
480 uncertainty declined with sample size, asymptoting at 250-1,000 individuals per species or per  
481 region (Appendix E). This suggests that future data collection activities should focus on species  
482 and regions with data densities below that threshold.

483

#### 484 **4.4 Challenges and opportunities**

485

486 In this study, predictive capacity for the models was moderate, as evidenced by the fit of the  
487 subset of observations with relatively small sampling error to the predicted phenological dates

488 (Fig 1). Consistent with the literature, our models performed better for spring than fall events  
489 (Gallinat et al., 2015). This may reflect a higher degree of variability in the drivers of onset of  
490 colored leaves such as disease or other stressors. Future applications using the analytical  
491 framework outline here might consider alternative or novel methods for accumulating  
492 phenological forcing variables. For example, modeling forcing variables using critical thresholds  
493 rather than using a simple logit-linear relationship; accounting for the varying importance of  
494 environmental drivers over different time periods, and/or modeling interactive effects between  
495 drivers may yield improved results. An additional consideration is that our estimates of  
496 sensitivity to environmental variation combine both sensitivity to interannual climate variability  
497 and sensitivity to climate variability over space (due to different locations being sampled in  
498 different years). While it is relatively common to combine shorter phenological time series in this  
499 way (Liang and Schwartz, 2014; Melaas et al., 2016; Taylor et al., 2019), even within regions  
500 there is some evidence that sensitivity to temperature over spatial gradients are stronger than  
501 over time (Jocher et al., 2013). As more years of data accumulate from the core-resampled  
502 (NEON) datasets here, it may be possible to address the generality of this pattern, and how to  
503 best model both spatial and temporal sensitivity. Approaches that account for the relationship  
504 between the timing of early-season and later-season phenological events may result in  
505 increased performance (Keenan and Richardson, 2015). On the other hand, a similar study  
506 (using probit-regression, which has a fairly similar sigmoid form relating linear terms of  
507 probability of occurrence as logit-regression) yielded very accurate predictions of flowering date  
508 (Allen et al., 2014). This suggests that some of the 'noise' inherent in many distributed  
509 phenology datasets, and/or generalizing over such large regions and taxonomically diverse  
510 species may have predictive limits.

511  
512 A limitation of the framework used here is that models were slow to run. Re-running the models  
513 over a large combination of forcing variables\*periods\*functional forms, as has been done for

514 some other efforts (Basler, 2016; Park and Mazer, 2018), is too computationally expensive at  
515 present. A more tractable approach to including alternative forms of environmental drivers or  
516 accumulation windows might be to use a smaller subset of species and regions for intermodel  
517 comparison before refitting the most promising subset on the full dataset.

518

## 519 **5. CONCLUSIONS**

520

521 The development of phenological prediction tools requires advancing the field of phenology  
522 beyond directional assessment, such as evaluating whether plants generally leaf out earlier with  
523 warmer springs, to specific quantitative assessments, such as determining the number of days  
524 earlier a species of plant growing in a particular location will leaf out with warming that occurs at  
525 particular times of year (Dietze et al., 2018; Houlahan et al., 2017). We demonstrated a novel  
526 approach to predicting the timing of phenological events such as leaf out utilizing survival  
527 analysis. This hierarchical approach demonstrated strengths over traditional statistical  
528 approaches, allowing for the incorporation of data across species and regions and leveraging  
529 the large amount of data availability while also offering information on species and region level  
530 variability. The results of this study offer a path forward for making widespread phenological  
531 predictions by moving from the development of individual models tailored to a single site and  
532 taxon to a generalized framework for predicting the transition states for any species, in any  
533 location.

534

## 535 **6. ACKNOWLEDGEMENTS**

536

537 We are thankful for thoughtful input from Alyssa Rosemartin, Ellen Denny, Katherine Jones, Lee  
538 Marsh and Jeff Switzer and computing support from Nolan Kane. Erynn Maynard and two  
539 anonymous reviewers provided helpful comments on the manuscript. Data were provided by the

540 USA National Phenology Network and the many participants who contribute to its *Nature's*  
541 *Notebook* program as well as the National Ecological Observatory and its many field  
542 technicians. The National Ecological Observatory Network is a program sponsored by the  
543 National Science Foundation and operated under cooperative agreement by Battelle Memorial  
544 Institute. This material is based in part upon work supported by the National Science Foundation  
545 through NEON and the USGS through USA-NPN. Any use of trade, firm, or product names is  
546 for descriptive purposes only and does not imply endorsement by the U.S. government.

547

548 **FUNDING**

549

550 This work was supported by Cooperative Agreements G14AC00405, G17AC00405, and  
551 G18AC00135 from the U.S. Geological Survey..

552

553

554

555

556 **REFERENCES**

557 Abramoff, R.Z., and Finzi, A.C. (2015). Are above- and below-ground phenology in sync? *New*  
558 *Phytol.* *205*, 1054–1061.

559 Allen, J.M., Terres, M.A., Katsuki, T., Iwamoto, K., Kobori, H., Higuchi, H., Primack, R.B.,  
560 Wilson, A.M., Gelfand, A., and Silander, J.A. (2014). Modeling daily flowering probabilities:  
561 expected impact of climate change on Japanese cherry phenology. *Glob. Change Biol.* *20*,  
562 1251–1263.

563 Archetti, M., Richardson, A.D., O’Keefe, J., and Delpierre, N. (2013). Predicting Climate Change  
564 Impacts on the Amount and Duration of Autumn Colors in a New England Forest. *PLOS ONE* *8*,  
565 e57373.

566 Basler, D. (2016). Evaluating phenological models for the prediction of leaf-out dates in six  
567 temperate tree species across central Europe. *Agric. For. Meteorol.* *217*, 10–21.

568 Bolker, B. M. (2008). *Ecological Models and Data in R*. Princeton University Press, Princeton,  
569 New Jersey.

570

571 Clark, J. S. 2007. *Models for ecological data: an introduction*. Princeton University Press,  
572 Princeton, New Jersey.

573 Cook, B.I., Wolkovich, E.M., and Parmesan, C. (2012). Divergent responses to spring and  
574 winter warming drive community level flowering trends. *Proc. Natl. Acad. Sci.* *109*, 9000–9005.

575 Cox, D.R. (1972) Regression models and life-tables. *J Royal Stat Soc Ser B* *34*:187–220.

576

577

578 Crimmins, T.M., Crimmins, M.A., and Bertelsen, C.D. (2010). Complex responses to climate  
579 drivers in onset of spring flowering across a semi-arid elevation gradient. *J. Ecol.* **98**, 1042–  
580 1051.

581 Denny, E.G., Gerst, K.L., Miller-Rushing, A.J., Tierney, G.L., Crimmins, T.M., Enquist, C.A.F.,  
582 Guertin, P., Rosemartin, A.H., Schwartz, M.D., Thomas, K.A., et al. (2014). Standardized  
583 phenology monitoring methods to track plant and animal activity for science and resource  
584 management applications. *Int. J. Biometeorol.* **58**, 591–601.

585 Dietze, M.C., Fox, A., Beck-Johnson, L.M., Betancourt, J.L., Hooten, M.B., Jarnevich, C.S.,  
586 Keitt, T.H., Kenney, M.A., Laney, C.M., Larsen, L.G., et al. (2018). Iterative near-term ecological  
587 forecasting: Needs, opportunities, and challenges. *Proc. Natl. Acad. Sci.* **115**, 1424–1432.

588 Diez, J.M., Ibáñez, I., Silander Jr, J.A., Primack, R., Higuchi, H., Kobori, H., Sen, A., and James,  
589 T.Y. (2014). Beyond seasonal climate: statistical estimation of phenological responses to  
590 weather. *Ecol. Appl.* **24**, 1793–1802.

591 Dorigo, W.A., Wagner, W., Albergel, C., Albrecht, F., Balsamo, G., Brocca, L., Chung, D., Ertl,  
592 M., Forkel, M., Gruber, A., et al. (2017) ESA CCI Soil Moisture for improved Earth system  
593 understanding: State-of the art and future directions, *Rem. Sens. of Envi.* **203**: 185-215.

594  
595 Elmendorf, S.C., Jones, K.D., Cook, B.I., Diez, J.M., Enquist, C.A.F., Hufft, R.A., Jones, M.O.,  
596 Mazer, S.J., Miller-Rushing, A.J., Moore, D.J.P., et al. (2016). The plant phenology monitoring  
597 design for The National Ecological Observatory Network. *Ecosphere* **7**, n/a-n/a.

598 Epstein, E.S. (1969). A Scoring System for Probability Forecasts of Ranked Categories. *J. Appl.*  
599 *Meteorol.* **8**, 985–987.

600 Fitchett, J.M., Grab, S.W., and Thompson, D.I. (2015). Plant phenology and climate change:  
601 Progress in methodological approaches and application. *Prog. Phys. Geogr. Earth Environ.* **39**,  
602 460–482.

603 Flynn, D.F.B., and Wolkovich, E.M. (2018). Temperature and photoperiod drive spring  
604 phenology across all species in a temperate forest community. *New Phytol.* **219**, 1353–1362.

605 Fu, Y.H., Piao, S., Delpierre, N., Hao, F., Hänninen, H., Liu, Y., Sun, W., Janssens, I.A., and  
606 Campioli, M. (2018). Larger temperature response of autumn leaf senescence than spring leaf-  
607 out phenology. *Glob. Change Biol.* **24**, 2159–2168.

608 Gallinat, A.S., Primack, R.B., and Wagner, D.L. (2015). Autumn, the neglected season in  
609 climate change research. *Trends Ecol. Evol.* **30**, 169–176.

610 Gelman, A., and Hill, J. (2007). *Data Analysis Using Regression and Multilevel/Hierarchical*  
611 *Models* (Cambridge University Press).

612 Gill, A.L., Gallinat, A.S., Sanders-DeMott, R., Rigden, A.J., Short Gianotti, D.J., Mantooth, J.A.,  
613 and Templer, P.H. (2015). Changes in autumn senescence in northern hemisphere deciduous  
614 trees: a meta-analysis of autumn phenology studies. *Ann. Bot.* **116**, 875–888.

615 Gunderson, C.A., Edwards, N.T., Walker, A.V., O'Hara, K.H., Campion, C.M., and Hanson, P.J.  
616 (2012). Forest phenology and a warmer climate - growing season extension in relation to  
617 climatic provenance. *Glob. Change Biol.* **18**, 2008–2025.

618 Houlahan, J.E., McKinney, S.T., Anderson, T.M., and McGill, B.J. (2017). The priority of  
619 prediction in ecological understanding. *Oikos* **126**, 1–7.

620 Hufft, R.A., DePrenger-Levin, M.E., Levy, R.A., and Islam, M.B. (2018). Using herbarium  
621 specimens to select indicator species for climate change monitoring. *Biodivers. Conserv.* 27,  
622 1487–1501.

623 Inouye, D.W. (2000). The ecological and evolutionary significance of frost in the context of  
624 climate change. *Ecol. Lett.* 3, 457–463.

625 Jochner, S., Caffarra, A., and Menzel, A. (2013). Can spatial data substitute temporal data in  
626 phenological modelling? A survey using birch flowering. *Tree Physiol* 33, 1256–1268.

627 Keenan, T.F., and Richardson, A.D. (2015). The timing of autumn senescence is affected by the  
628 timing of spring phenology: implications for predictive models. *Glob. Change Biol.* 21, 2634–  
629 2641.

630 Korner, C., and Basler, D. (2010). Phenology under global warming. *Science* 327, 1461–1462.

631 Lechowicz, M.J. (1984). Why do temperate deciduous trees leaf out at different times?  
632 Adaptation and ecology of forest communities. *Am. Nat.* 124, 821–842.

633 Liang, L. (2019). A spatially explicit modeling analysis of adaptive variation in temperate tree  
634 phenology. *Agric. For. Meteorol.* 266–267, 73-86.

635 Liang, L., and Schwartz, M.D. (2014). Testing a growth efficiency hypothesis with continental-  
636 scale phenological variations of common and cloned plants. *Int. J. Biometeorol.* 58, 1789–1797.

637 Ma, Q., Huang, J.-G., Hänninen, H., and Berninger, F. (2018). Reduced geographical variability  
638 in spring phenology of temperate trees with recent warming. *Agric. For. Meteorol.* 256–257,  
639 526–533.

640 MacKenzie, C.M., Primack, R.B., and Miller-Rushing, A.J. (2018). Local environment, not local  
641 adaptation, drives leaf-out phenology in common gardens along an elevational gradient in  
642 Acadia National Park, Maine. *Am. J. Bot.* 105, 986–995.

643 Mazer, S.J., Gerst, K.L., Matthews, E.R., and Evenden, A. (2015). Species-specific phenological  
644 responses to winter temperature and precipitation in a water-limited ecosystem. *Ecosphere* 6,  
645 art98.

646 McKinney, A.M., CaraDonna, P.J., Inouye, D.W., Barr, B., Bertelsen, C.D., and Waser, N.M.  
647 (2012). Asynchronous changes in phenology of migrating Broad-tailed Hummingbirds and their  
648 early-season nectar resources. *Ecology* 93, 1987–1993.

649 Melaas, E.K., Friedl, M.A., and Richardson, A.D. (2016). Multiscale modeling of spring  
650 phenology across Deciduous Forests in the Eastern United States. *Glob. Change Biol.* 22, 792–  
651 805.

652 Menzel, A. (2002). Phenology: Its Importance to the Global Change Community. *Clim. Change*  
653 54, 379–385.

654 Miller-Rushing, A.J., and Primack, R.B. (2008). Global warming and flowering times in  
655 Thoreau's Concord: A community perspective. *Ecology* 89, 332–341.

656 Morisette, J.T., Richardson, A.D., Knapp, A.K., Fisher, J.I., Graham, E.A., Abatzoglou, J.,  
657 Wilson, B.E., Breshears, D.D., Henebry, G.M., Hanes, J.M., et al. (2009). Tracking the rhythm of  
658 the seasons in the face of global change: phenological research in the 21st century. *Front. Ecol.*  
659 *Environ.* 7, 253–260.

660 Murphy, A.H. (1970). The ranked probability score and the probability score: a comparison.  
661 *Mon. Weather Rev.* 98, 917–924.

662 [dataset] National Ecological Observatory Network. 2018. Data Product: NEON.DP1.10055.  
663 Provisional data downloaded from <http://data.neonscience.org> on 2018-07-05. Battelle, Boulder,  
664 CO, USA.

665

666 Oberbauer, S., Elmendorf, S., Troxler, T., Hollister, R., Rocha, A., Bret-Harte, M., Dawes, M.,  
667 Fosaa, A., Henry, G., and Høye, T.T. (2013). Phenological response of tundra plants to  
668 background climate variation tested using the International Tundra Experiment. *Philos. Trans.*  
669 *R. Soc. B Biol. Sci.* **368**, 20120481.

670 Pan, F., Peters-Lidard, C.D., and Sale, M.J. (2003). An analytical method for predicting surface  
671 soil moisture from rainfall observations. *Water Resour. Res.* **39**.

672 Park, I.W., and Mazer, S.J. (2018). Overlooked climate parameters best predict flowering onset:  
673 Assessing phenological models using the elastic net. *Glob. Change Biol.* **24**, 5972–5984.

674 Parker, T.C., Tang, J., Clark, M.B., Moody, M.M., and Fetcher, N. (2017). Ecotypic differences  
675 in the phenology of the tundra species *Eriophorum vaginatum* reflect sites of origin. *Ecol. Evol.*  
676 **7**, 9775–9786.

677 Pearse, W.D., Davis, C.C., Inouye, D.W., Primack, R.B., and Davies, T.J. (2017). A statistical  
678 estimator for determining the limits of contemporary and historic phenology. *Nature Ecology &*  
679 *Evolution* **1**, 1876-1882.

680 Peng, D., Wu, C., Li, C., Zhang, X., Liu, Z., Ye, H., Luo, S., Liu, X., Hu, Y., and Fang, B. (2017).  
681 Spring green-up phenology products derived from MODIS NDVI and EVI: Intercomparison,  
682 interpretation and validation using National Phenology Network and AmeriFlux observations.  
683 *Ecological Indicators* **77**, 323–336.

684

685  
686 Piao, S., Tan, J., Chen, A., Fu, Y.H., Ciais, P., Liu, Q., Janssens, I.A., Vicca, S., Zeng, Z.,  
687 Jeong, S.-J., et al. (2015). Leaf onset in the northern hemisphere triggered by daytime  
688 temperature. *Nature Communications* 6, 6911.

689  
690 Prev y, J., Vellend, M., R ger, N., Hollister, R.D., Bjorkman, A.D., Myers-Smith, I.H., Elmendorf,  
691 S.C., Clark, K., Cooper, E.J., Elberling, B., et al. (2017). Greater temperature sensitivity of plant  
692 phenology at colder sites: implications for convergence across northern latitudes. *Glob. Change*  
693 *Biol.* 23, 2660–2671.

694 Reichle, R., G. De Lannoy, R. D. Koster, W. T. Crow, J. S. Kimball, and Q. Liu. 2018. *SMAP L4*  
695 *Global 3-hourly 9 km EASE-Grid Surface and Root Zone Soil Moisture Analysis Update, Version*  
696 *4*. Boulder, Colorado USA. NASA National Snow and Ice Data Center Distributed Active Archive  
697 Center. doi: <https://doi.org/10.5067/60HB8VIP2T8W>.

698

699 Richardson, A.D., Keenan, T.F., Migliavacca, M., Ryu, Y., Sonnentag, O., and Toomey, M.  
700 (2013). Climate change, phenology, and phenological control of vegetation feedbacks to the  
701 climate system. *Agric. For. Meteorol.* 169, 156–173.

702 Richardson, E.A., Seeley, S.D., and Walker, D.R. (1974). A model for estimating the completion  
703 of rest for “Redhaven” and “Elberta” peach trees. *HortScience* 9, 331-332.

704 Roberts, A.M.I., Tansey, C., Smithers, R.J., and Phillimore, A.B. (2015). Predicting a change in  
705 the order of spring phenology in temperate forests. *Glob Change Biol* 21, 2603–2611.

706 Royle, J.A., and Dorazio, R.M. (2008). Modeling Survival. In *Hierarchical Modeling and*  
707 *Inference in Ecology: The Analysis of Data from Populations, Metapopulations and*  
708 *Communities*, (Academic Press), pp. 347- 377.

709 Siljamo, P., Sofiev, M., Filatova, E., Grewling, Ł., Jäger, S., Khoreva, E., Linkosalo, T., Ortega  
710 Jimenez, S., Ranta, H., Rantio-Lehtimäki, A., et al. (2013). A numerical model of birch pollen  
711 emission and dispersion in the atmosphere. Model evaluation and sensitivity analysis. *Int. J.*  
712 *Biometeorol.* *57*, 125–136.

713 Taylor, S.D., Meiners, J.M., Riemer, K., Orr, M.C., and White, E.P. (2019). Comparison of large-  
714 scale citizen science data and long-term study data for phenology modeling. *Ecology* *100*,  
715 e02568.

716 Templ, B., Fleck, S., and Templ, M. (2017). Change of plant phenophases explained by survival  
717 modeling. *Int. J. Biometeorol.* *61*, 881–889.

718 Terres, M.A., Gelfand, A.E., Allen, J.M., and Silander, J.A. (2013). Analyzing first flowering  
719 event data using survival models with space and time-varying covariates. *Environmetrics* *24*,  
720 317–331.

721 [dataset] Thornton, P.E., Thornton, M.M., Mayer, B.W., Wei, Y., Devarakonda, R., Vose, R.S.,  
722 and Cook, R.B. (2018). Daymet: Daily Surface Weather Data on a 1-km Grid for North America,  
723 Version 3. Data set accessed 5 July 2018. ORNL DAAC Oak Ridge Tenn. USA.  
724 <https://doi.org/10.3334/ORNLDAAC/1328>

725  
726 [dataset] United States Environmental Protection Agency. 2018. Ecoregions of North America.  
727 USA. Data set accessed 2018-07-05 at [https://www.epa.gov/eco-research/ecoregions-north-](https://www.epa.gov/eco-research/ecoregions-north-america)  
728 [america](https://www.epa.gov/eco-research/ecoregions-north-america)

729  
730 [dataset] USA National Phenology Network. 2018. Plant and Animal Phenology Data. Data type:  
731 Status and Intensity (2009-2017). NPN, Tucson, Arizona, USA. Data set accessed 2018-07-05  
732 at <http://doi.org/10.5066/F78S4N1V>

733

734 Way, D.A., and Montgomery, R.A. (2015). Photoperiod constraints on tree phenology,  
735 performance and migration in a warming world. *Plant Cell Environ.* **38**, 1725–1736.

736 Williamson, S.N., Barrio, I.C., Hik, D.S., and Gamon, J.A. (2016). Phenology and species  
737 determine growing-season albedo increase at the altitudinal limit of shrub growth in the sub-  
738 Arctic. *Glob. Change Biol.* **22**, 3621–3631.

739 Xie, Y., Ahmed, K.F., Allen, J.M., Wilson, A.M., and Silander, J.A. (2015). Green-up of  
740 deciduous forest communities of northeastern North America in response to climate variation  
741 and climate change. *Landsc. Ecol.* **30**, 109–123.

742 Xie, Y., Civco, D.L., and Silander, J.A. (2018). Species-specific spring and autumn leaf  
743 phenology captured by time-lapse digital cameras. *Ecosphere* **9**, e02089.

744 Zhang, X., Liu, L., Liu, Y., Jayavelu, S., Wang, J., Moon, M., Henebry, G.M., Friedl, M.A., and  
745 Schaaf, C.B. (2018). Generation and evaluation of the VIIRS land surface phenology product.  
746 *Remote Sensing of Environment* **216**, 212–229.

747

748

749  
750  
751  
752  
753  
754  
755  
756  
757  
758  
759  
760  
761  
762  
763  
764  
765  
766  
767  
768  
769  
770  
771  
772  
773

**SUPPLEMENTAL TABLES**

Table S.1: Species-specific parameter estimates for the spring leaf out model. Shown are the sensitivity of plants in each taxon to each forcing variable (+ intercept). Included are point estimates + 95% CI for each coefficient  $\beta_{j,species}$ . Random effects for species are  $u_j$  centered (see appendix A, Statistical modeling details); negative parameter estimates reflect species where higher values of the coefficient accelerates spring leaf out. See figure 3 in the main text for an exemplar graphical display of the same information for  $\beta_{GDD,species}$ .

Table S.2: Region-specific parameter estimates for the spring leaf out model. Shown are the sensitivity of plants in each region to each forcing variable (+ intercept). Included are point estimates + 95% CI for each coefficient  $\beta_{j,region}$ . Random effects for region are 0-centered (see appendix A, Statistical modeling details). Negative parameter estimates therefore reflect regions where higher values of the coefficient accelerates spring leaf out more *relative to other regions*.

Table S.3: Species-specific parameter estimates for the fall leaf-coloration model. Shown are the sensitivity of plants in each taxon to each forcing variable. Included are point estimates + 95% CI for each coefficient  $\beta_{j,species}$ . Random effects for species are  $u_j$  centered (see appendix A, Statistical modeling details); negative parameter estimates reflect species where higher values of the coefficient accelerate fall leaf coloration leaf out.

774 Table S.4: Region-specific parameter estimates for the fall leaf-coloration model. Shown are the  
775 sensitivity of plants in each region to each forcing variable. Included are point estimates + 95%  
776 CI for each coefficient  $\beta_{j,region}$ . Random effects for region are 0-centered (see appendix A,  
777 Statistical modeling details). Negative parameter estimates therefore reflect regions where  
778 higher values of the coefficient accelerate fall leaf coloration more *relative to other regions*.  
779  
780  
781