




# Genome Sequences of *Allochrochromatium palmeri* and *Allochrochromatium humboldtianum* Expand the *Allochrochromatium* Family Tree of Purple Sulfur Photosynthetic Bacteria within the *Gammaproteobacteria* and Further Refine the Genus

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**ABSTRACT** New genomes of two *Allochrochromatium* strains were sequenced. Whole-genome and average nucleotide identity based on BLAST (ANIb) comparisons show that *Allochrochromatium humboldtianum* is the nearest relative of *Allochrochromatium vinosum* (ANIb, 91.5%), while both *Allochrochromatium palmeri* and *Thiochromatium tepidum* are more distantly related (ANIb, <87%). These new sequences firmly establish the position of *Allochrochromatium* on the family tree.

*Chrochromatium vinosum* (now *Allochrochromatium vinosum*) is the prototypic purple sulfur bacterium, and it is the only species in the genus to have had a genome sequence determined (1). Moreover, there are several genera that are fairly closely related to *Allochrochromatium*, including *Thiocystis*, *Thiochromatium*, *Chrochromatium*, and *Thiorhodococcus* (2), although the relationships are not clear despite single-gene comparisons (3); therefore, a whole-genome comparison including multiple *Allochrochromatium* species is needed.

*Allochrochromatium palmeri* DSM 15591<sup>T</sup> was originally isolated from a cave system in the Bahamas (4), while *Allochrochromatium humboldtianum* DSM 21881<sup>T</sup> was isolated from marine sediments in Peru (5). Cultures were grown and genomic DNA was prepared by the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ). DNA analysis showed  $A_{260}/A_{280}$  ratios of 1.60 for *A. palmeri* and 1.96 for *A. humboldtianum*. The sequencing libraries were prepared using the Illumina Nextera DNA Flex library preparation kit and were sequenced by an Illumina MiniSeq sequencer using 500  $\mu$ l of a 1.8 pM library. Paired-end (2  $\times$  150-bp) sequencing generated 2,433,982 reads and 192 Mbp for *A. palmeri* and 3,349,346 reads and 252.2 Mbp for *A. humboldtianum*. Quality control of the reads was performed using FastQC within BaseSpace (version 1.0.0; Illumina), using a k-mer size of 5 and contamination filtering. We assembled the genome *de novo* through PATRIC (6) using SPAdes (version 3.10.0) (7) for *A. palmeri* and Unicycler for *A. humboldtianum*. The assembly yielded 196 contigs (>300 bp) and an  $N_{50}$  value of 74,142 bp for *A. palmeri* (45 $\times$  coverage), while *A. humboldtianum* was assembled into 86 contigs with an  $N_{50}$  value of 305,111 bp (55 $\times$  coverage). The *A. palmeri* genome had a GC content of 62.5% and a length of 4,272,782 bp, whereas the *A. humboldtianum* genome had a GC content of 63.9% and a length of 4,584,820 bp. The genomes were annotated using the RAST tool kit (RASTtk) (8) within PATRIC (6). This annotation showed *A. palmeri* to have 4,134 coding sequences and 45 tRNAs and *A. humboldtianum* to contain 4,391 coding sequences and 47 tRNAs. Default parameters were used for all software applications unless otherwise noted.

A JSpeciesWS comparison (9) of average nucleotide identity based on BLAST (ANIb)

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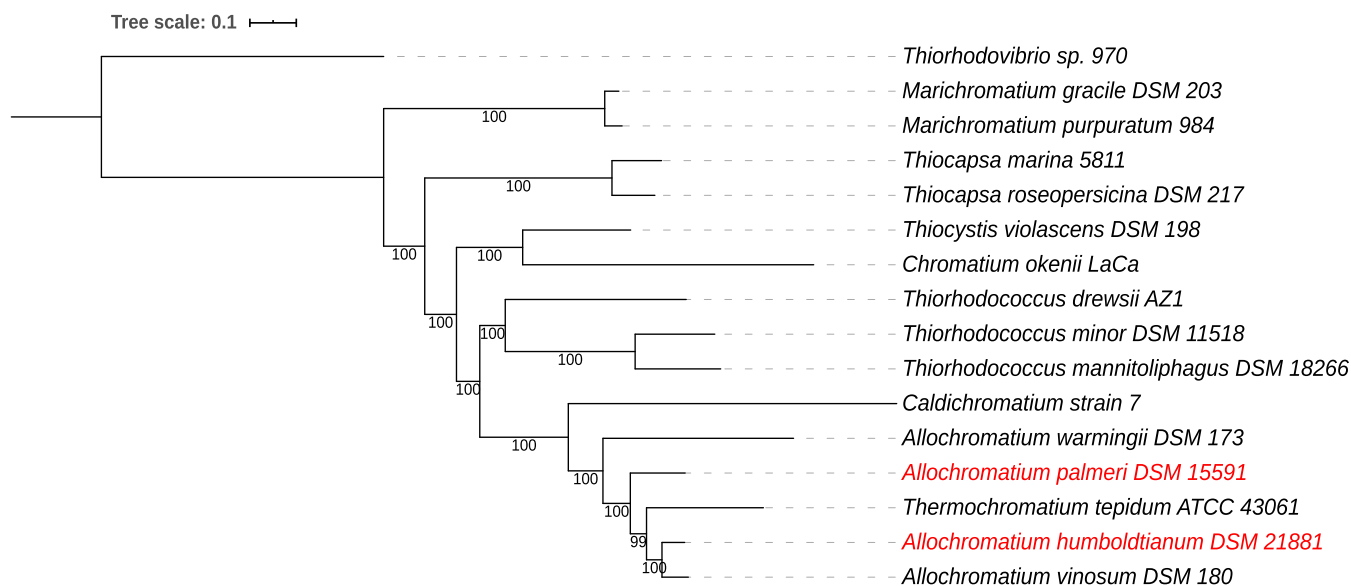
**TABLE 1** ANIb comparisons

Strain	ANIb (%) with strain:			
	<i>A. vinosum</i> DSM 180 <sup>T</sup>	<i>A. humboldtianum</i> DSM 21881 <sup>T</sup>	<i>A. palmeri</i> DSM 15591 <sup>T</sup>	<i>T. tepidum</i> ATCC 43061 <sup>T</sup>
<i>A. humboldtianum</i> DSM 21881 <sup>T</sup>	91.5			
<i>A. palmeri</i> DSM 15591 <sup>T</sup>	86.6	86.8		
<i>T. tepidum</i> ATCC 43061 <sup>T</sup>	84.3	84.9	82.2	
<i>A. warmingii</i> DSM 173 <sup>T</sup>	76.6	76.5	76.4	74.7

showed 86.8% identity between *A. palmeri* and *A. humboldtianum* (Table 1). *A. humboldtianum* is closer to *Allochrochromatium vinosum* with 91.5% ANIb, while *A. palmeri* showed 86.6% ANIb. All of these ANIb values are clearly below the proposed 95% cutoff value for genome definition of a species (9). *Thermochromatium tepidum* is about equidistant from all three of the *Allochrochromatium* species; however, *Allochrochromatium warmingii* appears to be more distant from all of them.

Whole-genome-based phylogenetic analysis was performed with RAxML within PATRIC (10, 11) using all of the *Allochrochromatium* and related genomes (1, 12–16). This analysis grouped all of the *Allochrochromatium* species (Fig. 1); however, it also placed *Thermochromatium tepidum* within this group. Consistent with the ANIb analysis, *A. warmingii* is more distant from the other *Allochrochromatium* species. Further genetic and physiological studies may be needed to determine whether a nomenclature change of the latter species is warranted. The addition of these new *Allochrochromatium* genomes has substantially strengthened the phylogenetic tree of this genus.

**Data availability.** These whole-genome shotgun projects have been deposited in DDBJ/ENA/GenBank under the accession numbers [WNKT000000000](https://www.ncbi.nlm.nih.gov/nuccore/WNK000000000) for *Allochrochromatium palmeri* and [JABZEO000000000](https://www.ncbi.nlm.nih.gov/nuccore/JABZEO000000000) for *Allochrochromatium humboldtianum*. The versions described in this paper are versions [WNKT010000000](https://www.ncbi.nlm.nih.gov/nuccore/WNK010000000) and [JABZEO010000000](https://www.ncbi.nlm.nih.gov/nuccore/JABZEO010000000). The raw sequencing reads have been submitted to SRA, and the accession numbers are [SRR12110462](https://www.ncbi.nlm.nih.gov/sra/SRR12110462) for *Allochrochromatium palmeri* and [SRR12110432](https://www.ncbi.nlm.nih.gov/sra/SRR12110432) for *Allochrochromatium humboldtianum*.



**FIG 1** Whole-genome-based phylogenetic tree of all sequenced *Allochrochromatium* and related species. The phylogenetic tree was generated using the Codon Tree method within PATRIC (6), which used PATRIC global protein families (PGFams) as homology groups; 467 PGFams were found among these selected genomes using the Codon Tree analysis, and the aligned proteins and coding DNA from single-copy genes were used for RAxML analysis (10, 11). The support values for the phylogenetic tree are shown on the tree branches and were generated using 100 rounds of the rapid bootstrapping option of RAxML. *Thiorhodovibrio* was used as an outgroup. Interactive Tree Of Life (iTOL) was used for the tree visualization (17).

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